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The influence of different air-drying conditions on bioactive compounds and antioxidant activity of berries

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ABSTRACT

The aim of the present research was to study the effect of convective drying on color, bioactive compounds and antioxidant activity of berry fruits and to chemically characterize the polyphenolic composition of raspberry, boysenberry, redcurrants and blackcurrants fruit. Drying berries at 65 °C provoked the best conservations of color, particularly for boysenberry and blackcurrant. Drying at 65 °C was also the condition that showed higher level of polyphenols, while drying at 50 °C or 130 °C showed above % degradation of them due to the long time or high temperature drying. Radical scavenging activity was the predominant antioxidant mechanism in all samples, being 65 °C dried berries the most active ones possibly due to polyphenols depolymerization. The anthocyanin profile showed that delphinidin and cyanidin derivatives were the most abundant anthocyanidins with different predominance between berry genera. Degradation of anthocyanins was increased with drying temperature been Cy 3-glucoside and Cy 3-rutinoside the most abundant.

1.- INTRODUCTION

Berry fruits are a type of small fine fruits characterized by a red, purple and blue color. The most common are: blueberry, cranberry, blackberry, raspberry, white, red or blackcurrant and strawberry. The consumption of these fruits is mainly restricted to fresh product or processed food such as juice, beverage and jam due to their short shelf-life, so that different marketing strategies like drying and/or packaging can be used to retain the quality of berries during storage. In this sense, dried or frozen berries are the two most commonly consumed forms of these fruits despite the decrease in nutritional value found in the former or the high cost of transportation in the latter.

Berries contain high levels of polyphenols including flavonoids (anthocyanins, flavonols and flavanols such as condensed tannins or proanthocyanidins), hydrolysable tannins (ellagitannins and gallotannins), phenolic acids (hydroxybenzoic and hydroxycinnamic acids, chlorogenic acid), stilbenoids and lignans. Many studies have been published in relation to the health benefits of berry consumption due to the presence of those active compounds.

Thanks to the rich and diverse composition of bioactive compounds and their health-promoting properties which result mostly from their antioxidant activity, their metal chelating capacity and their affinity for proteins, berry fruits are widely recognized as natural functional products.

Since the availability of berries is seasonal, their processing for long-term storage and their addition to the formulation of new products are a good way to improve the nutritional quality of final products. The most commonly used methods of food preservation involve thermal treatments to reduce moisture content to a safe level, depriving...
molds of favorable proliferative conditions\textsuperscript{9,10}. In this sense, convective drying is always an alternative to extend shelf life and allow powders to be used as additives of other food matrixes, promoting the use of these types of highly nutritious fruits\textsuperscript{11,12}.

It should be noted that quality characteristics of dehydrated fruits will be affected by drying conditions, in which phytochemicals like anthocyanins and phenolic compounds are highly susceptible to degradation\textsuperscript{13}, thus reducing their antioxidant activity, and in general, their functional characteristics. Some studies related to drying berries indicate that this technological process may have negative effects on bioactive compounds compared to their fresh counterpart\textsuperscript{11}. Therefore, the conditions of the drying process can produce light or moderate effects on fruit's bioactive compounds, mainly depending on the type of drying process and the fruit species\textsuperscript{14}. As a result, dried fruits could not only be a good source of vitamins and fiber but also provide a wide array of bioactive phytochemicals that have been linked to a reduction in the risk of chronic diseases. Profiling the bioactive compounds that remains in dried fruits is the first step to address benefits associated to their consumption.

Since drying process affects appearance and chemical composition, the best method should be selected to ensure maximum conservation of bioactive compounds. Freeze drying is probably the best technique to reach high quality dried fruits; however, it is also one of the most expensive ones\textsuperscript{15}. Moreover, freeze drying is the best method for obtaining a high-quality dried product (for heat sensitive materials) since it is commonly considered that minor modifications take place\textsuperscript{16}. On the other hand, convective drying is the most economical and widely adopted technique in the food industry, despite requiring long drying times and high temperatures\textsuperscript{11}. In the last years, new emerging technologies for drying fruits have been studied\textsuperscript{9}, although due to fine fruits like berries are commonly more expensive than others, their applications increases the cost even more. This fact reinforces the study of traditional drying technologies in order to determine the best conditions to preserve the characteristics of dried fruits that could be used to food enrichment without considerable costs increase of final product. Additionally, the effects of drying on polyphenolics and antioxidant activities have not been systematically studied, this selecting the best conditions to maximize presentation of color, anthocyanins and antioxidant activity, is crucial to produce dried fruit powder to be used as food coloring, snack production or functional ingredients to be included in other food products. Therefore, the aim of this work was to study the impact of convective drying at different temperatures those characterizes of berry fruits and to chemically characterize their polyphenolic composition.

\textbf{2.- MATERIALS AND METHODS}

\textbf{2.1.- Materials}
The berry fruits for the present research were selected from four species, each two belonging to a different genus. Raspberry (*Rubus idaeus* var. Autumn Bliss) and boysenberry (*R. ursinus × R. idaeus* var. Black Satin) were chosen from *Rubus* genus, and redcurrants (*Ribes rubrum* sp.) and blackcurrants (*Ribes nigrum* sp.) from *Ribes* genus. All berries were purchased from Dolphes Gourmet (Rosario, Argentina) as Individual Quick Frozen (IQF) fruits, assuring low-quality changes and longer conservation times. *Rubus* genus fruits came from San Pedro, Buenos Aires (Argentina) while *Ribes* genus fruits from El Bolsón, Río Negro (Argentina). Fruits from two different crop years were purchased and combined in two pools considered as duplicates (raspberry and boysenberry from 2014-2015 and redcurrants and blackcurrants from 2013-2014).

All chemicals were of analytical grade unless otherwise stated. Formic and clorhidric acids, methanol and acetonitrile (HPLC grade) were obtained from Fisher Scientific (Madrid, Spain). Delphinidin-3-glucoside, cyanidin-3-glucoside, peonidin-3-glucoside, pelargonidin-3-glucoside and malvidin-3-glucoside standards were purchased from Extrasynthese (Lyon, France).

### 2.2.- Methods

**2.2.1.- Physicochemical characteristics of selected berry fruits**

Water content (method 934.06), ash (method 930.35) and proteins (method 920.152) were determined according to AOAC methods. It should be noted that fruits and their seeds were analyzed.

**2.2.2.- Berries drying**

Figure 1 shows the procedure used for drying berries. Briefly, each berry sample was separated in three sets in order to apply different convective drying conditions: 50 °C for 48 h, 65 °C for 20 h or 130 °C for 2 h until a moisture content below 15% was obtained. Until that, berries were blanched for 3 min in a home steamer (Smart-Tek SD2071, Argentina) for better conservation of anthocyanins and polyphenols and inactivation of polyphenol oxidase from berries as reported by Sablani *et al.*,¹⁵. The blanched fruits were carefully placed in a plastic mesh inside the air convection drier and separated in three portions and were exposed to the previously described conditions. Additionally, freeze-dried berries were obtained by storage at -80 °C and freeze dried for 48 h (L-T8 RIFICOR, Argentina). These samples were used as a reference assuming no significant modifications induced by lyophilization method. All samples were ground in a coffee mill (PE-MC9100, Peabody, Argentina) and stored at -18 °C prior to analysis. Water activity was measured by using an electric hygrometer Novasina Lab MASTER-aw (Novasina AG,
Lanchen, Switzerland) at 25°C. Measurements were done in duplicate and are expressed as the media ± standard deviation.

2.2.3.- Color of dried berries

The color of dried fruit powders was measured with a spectrophotometer (Minolta, Ramsey, NJ). Eight-millimeter measurement apertures, D65 illuminant, 10° angle of observer was settled and color recorded in CIE Lab space. L* indicates lightness, its value ranging from 0 (black) to 100 (white); a* and b* are the chromaticity coordinates. From the CIELAB coordinates, color function chroma ($\Delta C_{ab}$) and total color change ($\Delta E$) were calculated according to the following equations:

$$\Delta C_{ab} = (\Delta a^* + \Delta b^*)^{0.5}$$

$$\Delta E = (\Delta L^* + \Delta a^* + \Delta b^*)^{0.5}$$

2.2.4.- Bioactive compounds and antioxidant activity in dried berries

2.2.4.1.- Anthocyanin analysis

2.2.4.1.1.- Anthocyanin profile by HPLC-DAD-ESI/MS-QTOF

The analytical protocol for anthocyanins profile determination of the different samples studied is presented in Figure 1. Briefly, the method of anthocyanin purification was performed according to García-Herrera et al. A hydro-alcoholic extraction was done and samples were purified by an aqueous extraction followed by a amethanolic one, both extracts were mixed and passed through Agilent Tech cartridge (Figure 2). This extract was used to obtain the anthocyanin profile by HPLC analysis carried out on a liquid chromatography system (Hewlett Packard Agilent 1200 Series) equipped with a quaternary pump and a photo- diode array detector (DAD) (Agilent Technologies). The column used was a Phenomenex Luna C18 column (5 µm, 4.6 mm x 150 mm), set thermostatically at 25 °C. Chromatographic data were acquired and processed using an Agilent Chemstation for LC 3D system (Rev. B.04.01, Agilent Technologies). Briefly, the binary mobile phase used for analysis was aqueous 0.1% formic acid and HPLC-grade acetonitrile at a flow rate of 0.5 mL min$^{-1}$. Samples were analyzed in triplicate. Peaks were identified by comparing their retention time (Rt) and UV– visible spectra with the reference compounds, and the data were quantified using the corresponding curves of the reference compounds as standards, when available.

To confirm the identity of the compounds recorded, additional analyses were performed using HPLC coupled with mass spectrometry detection (HPLC-MS- TOF): liquid chromatography/mass selective Agilent 1200: quaternary pump (G1311A), diode array detector (G1315B). The column used was an Phenomenex Luna C18 column (5 µm, 4.6 mm x 150 mm); Mass QTOF (SAcetopectrometre Agilent G6530A Accurate Mass Q- TOF LCMS) with Electrospray
Ionization (ESI) with JetStream technology; Instrument State: standard dynamic range m/z 3200. Software used was MassHunter Data Acquisition B.04.00 and MassHunter Qualitative Analysis B.04.00.

2.2.4.1.2. Monomeric anthocyanin content

Four different solvent mixtures were prepared in order to extract the largest quantity of bioactive compounds from the whole fruits. Fifty milligrams of freeze-dried berries or 500 mg of air-dried were mixed with 1.5 ml of each solvent mixture: methanol: water (70:30), methanol: water (50:50), ethanol: water (70:30) and acetone: water (70:30) in all cases with a final concentration of 0.1% HCl. The solvent/berry mixtures were mixed for 10 min and then centrifuged at 12,000 g for 15 min. The supernatant was recovered and seven more extractions were performed. All determinations were made in the supernatants recovered. The solvent with better extraction performance based on the total anthocyanin content method was acetone: water (70:30) (data not shown), thus selected to determine total polyphenol content and antioxidant activity analysis (Figure 2).

Total anthocyanin content was performed according to Giusti and Wrolstad (2001) in the acidified dried berry extracts using an extinction coefficient (B) of 26,900 l cm$^{-1}$ mg$^{-1}$ and a molecular weight (MW) of 449.2 g/mol of cyanidin 3-glucoside. Absorbance were measured at 520 nm ($A_{520}$) and 700 nm ($A_{700}$) and calculated as follows:

$$\text{Cyanidin 3-glucoside } \mu g/g \text{ dried fruit} = \frac{((A_{520} - A_{700})_{pH\ 1^-} - (A_{520} - A_{700})_{pH4.5}) \times MW \times F \times Ve \times 10^5}{(B \times 1 \times Ws)},$$

F being the dilution factor for sample, Ve the extract volume and Ws the sample weight.

2.2.4.2.- Total polyphenol content

Total polyphenol content of dried berries was determined in two extracts as indicated in Figure 2, that was the methanolic extract made for anthocyanin profiling and in the acidified acetone extract for extraction of bioactive components previously described. Total polyphenols were determined using the Folin-Ciocalteu method, with gallic acid as a calibration standard. The concentration of total polyphenols was expressed as mg gallic acid per gram of fruit powder.

2.2.4.3.- Antioxidant activity determinations

2.2.4.3.1.- ABTS$^{•+}$ radical cation scavenging activity

The ABTS$^{•+}$ radical cation scavenging activity was measured according to Re et al. Trolox (Sigma 238813) was used as a standard and results were expressed as µmol of Trolox equivalent per gram of fruit powder.

2.2.4.2.3.- Ferric-reducing ability

Ferric reducing activity of dried fruits was determined by FRAP assay according to Pulido et al., using gallic acid as a standard.
2.2.5.- Statistical analysis

All samples were prepared in duplicate and each replicate was quantified in duplicate. Results were analyzed by the adjustment to a model with fixed effects for a classification factor with eight levels (drying methods and fruit type). The model included a variance function to take account of the presence of an increasing variability pattern related to medium levels of response variable. The adjustment was carried out using an implementation in InfoStat Software of gls function from the nlme library of R. The variance function applied was a function of implementation of power variance varPower() from nlme library. This type of statistical analysis allows comparing, simultaneously, the effect of drying method and fruit type. Results of the analyses were evaluated by using DGC test with a degree of significance of P<0.05. Pearson correlation coefficients were used to determine the relationship between anthocyanins contents determined by chromatographic and spectrophotometric methods.

2.3.- RESULTS AND DISCUSSION

2.3.1.- Physicochemical characteristics of selected berry fruits

The selected berry fruits were characterized according to their physicochemical composition as shown in Table 1. The four berry fruits showed high water content, raspberry being the one with the highest value (P<0.05). Blackcurrant presented the highest ash content, almost twice that of the observed for raspberry and boysenberry. Protein content was significantly higher for raspberry compared with both currant and boysenberry. Carbohydrate content was twice higher for blackcurrant compared with raspberry, while boysenberry and redcurrant showed intermediate values. USDA database was used as a reference for water content, protein and carbohydrate content of berries. Water content of selected berry fruits was very similar to the corresponding USDA value; yet, selected species showed increased protein content and slight difference in carbohydrate percentage compared to the standard value.

Water activity is a factor that highly influence dried fruit stability. A high water activity can lead to a shorter storage time of products, which is due to the possibility of microbial growth and biochemical changes, has been suggested a value bellow 0.600 to eliminate these factors. In this sense, the water activity of dried berries at different conditions ranged from 0.242 to 0.413 (Table 2). Considering each condition applied for drying fruits the final water activity decreased with increasing drying temperature, showing the following order: freeze-dried (0.3269)> 50 °C (0.3828)> 65 °C (0.3156)> 130 °C (0.2883). Water absorption values was very similar between all dried fruit within one specie, being on average: 0.309 for raspberry, 0.344 for boysenberry, 0.321 for redcurrant and 0.339 for
blackcurrant, similar values was found by Samoticha et al.,\textsuperscript{29} after convective drying of chokeberries. As a result, it can be assumed that the obtained dried berries powders were microbiologically stable.

\textbf{2.3.2. Color of dried berries}

Color and appearance in berries attract the consumer, they being associated with higher hedonic responses in fruits with darker color or higher anthocyanin content.\textsuperscript{1,30} Additionally, changes in color during thermal processing of fruit might provide information about alterations in the content of anthocyanins and other polyphenols. Statistical analysis showed that not only fruit type determined color variables but also drying condition influences them with a significant interaction in luminosity ($L^*$), redness ($a^*$) and yellowness ($b^*$).

Luminosity of dried fruits decreased with increasing drying temperature in all fruits, raspberry samples being those with highest $L^*$ values in all cases. In addition, drying blackcurrants at all convective drying conditions generated a large decrease in $L^*$ with no significant difference ($p>0.05$) between convective treatments (Figure 3). As seen in Figure 3, in addition to the significant interaction between fruit species and drying conditions, freeze drying was the best option to obtain dried fruits with high luminosity ($L^*= 34.46$ on average), followed by drying at 65 °C ($L^*= 29.52$ on average). On the other hand, convective drying at 130 °C decreased largely luminosity with a mean value of 24.31.

From Figure 3 it is clear that redness was the most affected color parameter during convective drying of berries. In addition, slight differences were detected between fruit drying at 50°C or 65°C ($a^*= 7.38$ on average) compared to freeze-drying ($a^*= 25.62$ on average) and high temperature drying ($a^*= 3.89$ on average). That means that the drying method showed a main effect on redness compared to fruit species. As observed with $L^*$ values, the highest redness was observed in freeze-dried berries. This could indicate that discoloration and browning during air drying may be the result of various chemical reactions including pigment destruction.\textsuperscript{31}

Since selected fruits are red/purple, yellowness was the less affected color variable; only drying at 130 °C showed a clear effect with the lowest $b^*$ values ($b^*= 1.81$ on average). Raspberry presented the highest yellowness in all drying conditions (except for freeze-dried conditions), while blackcurrant showed the minimum yellowness in all conditions. For \textit{Rubus} genus drying at 50 °C and 65 °C increased yellowness compared to freeze-dried conditions (Figure 3).

For better analysis of color modifications due to fruit species or drying conditions, chroma ($\Delta C_{ab}$) and total color difference ($\Delta E$) were determined (Table 3) using freeze-drying condition as a reference. Since chroma included variations in redness and yellowness, while total color difference also included luminosity, the similar values
observed in both parameters indicated that L* showed no considerable disparity between different dehydrated fruits species. Boysenberry showed the minimum chrome and total color differences in all conditions followed by blackcurrant, indicating that blue fruits better conserved color parameters, while major changes were found for raspberry dried at 130 °C.

2.3.3. Bioactive compounds and antioxidant activity in dried berries

2.3.3.1. Anthocyanin analysis

Anthocyanin quantification was carried out using two methods: HPLC-DAD-ESI/MS-QTOF (anthocyanin profile) and a spectrophotometric estimation (monomeric anthocyanins expressed as µg cyanidin 3-glucoside). Anthocyanin profile allows identifying different compounds in each fruit species as shown in Table 4, which is particularly new for boysenberry (blackberry hydride) selected in presented research and shown in Figure 4. In this sense, three different anthocyanin compounds were found in raspberry while four were detected in boysenberry, cyanidin 3-glucoside being found in both fruits. Analysis of redcurrant (RC) and blackcurrant (BC) (Ribes genus) showed only two compounds found in each one, being cyaniding 3-rutinoside the one in common.

Quantification of each identified anthocyanin was performed and results shown in Table 5. Analysis of anthocyanin profile showed that freeze-drying was the condition with the highest anthocyanin content in all samples. On the other hand, air-drying at 130 °C destroys anthocyanins, quantification in both purple fruits (boysenberry and blackcurrant) being minimum, in agreement with the low values in total color difference and chroma results described above.

Cy-3-sophoroside was co-eluted with Cy-3-sophoroside-5-rhamnoside in our conditions and thus quantified together. These two cyanidin compounds were the major anthocyanins in raspberry, reaching 52.3% of total anthocyanin in lyophilized form, in agreement with other authors. Compared with freeze-drying, only 56% and 25% of those anthocyanins were conserved when drying at 50 °C and 65 °C was applied, respectively, while no anthocyanin was detected in raspberry dehydrated at 130 °C. A similar proportion of Cy-3-glucoside conservation at intermediate drying temperatures was also detected for that fruit.

The anthocyanin profile of boysenberry was consistent with those of previously reports in blackberries with a single major peak representing Cy-3-glucoside that constituted 88.8% of the total anthocyanin content in the freeze-dried form. It should be noted that, in addition to decreasing anthocyanin content compared to freeze-drying conditions, dehydrating boysenberry by air-drying showed minor differences between 50°C and 65°C with 22% and
27% of Cy-3-glucoside conserved, respectively. In addition, in samples dried at 130°C, a minimum quantity of four anthocyanins was found. Delphinin-3-rutinoside and Cy-3-xylosyl-rutinoside were present in redcurrant at all drying conditions (except for drying at 130°C), Cy-3-xylosyl-rutinoside accounting for around 72.8% of total anthocyanins. As observed for boysenberry, drying at 50°C and 65°C produced the similar loss of compounds compared to freeze-dried redcurrant (Table 5). Blackcurrant showed equal quantity of rutinoside derivatives conserved after drying at 50°C compared to lyophilized form. Although no anthocyanin was found in blackcurrant dehydrated at 65 °C and a depreciable amount was found after drying at higher temperature, the profile observed in both Ribes genera was similar to those of other authors that reported delphinine-3-rutinose in both fruits with higher content in blackcurrant than in redcurrant. Figure 5 shows that results similar to the total anthocyanin detected by HPLC-DAD method were obtained when estimating the monomeric anthocyanin concentration via the spectrophotometric method with a Pearson’s correlation coefficient of 0.76 (P<0.05). In general, blackcurrant was the fruit with the highest anthocyanin content in all conditions (except drying at 65°C by HPLC-DAD method) followed by boysenberry, which could be attributed to the dark color of these berry species. Similar results were found by Arancibia-Avila, et al. by comparing raspberry and blueberries, indicating that color differences where closely associated to anthocyanins content. In this regard, raspberry and redcurrant showed the lowest anthocyanin content, showing similar values between them. With both drying methods a detrimental effect of drying conditions was observed, from which freeze-drying was found to be the method which allowed conservation of the highest concentration of anthocyanins. Drying berries at 50 °C or 65 °C produced fruits with similar anthocyanin content and the differences detected in freeze-dried berries according to each fruit was conserved but less pronounced. On the other hand, drying at 130 °C showed almost complete loss of anthocyanins (Figure 5).

In general, anthocyanin degradation seems to be more related to drying temperature than with exposure time since with increase in temperature, the time needed to dry fruits was reduced. As a result, anthocyanins were destroyed with increased drying temperature in a different proportion according to fruit species (significant interaction), in agreement with Karam et al. Drying berries at intermediate temperatures produced fruit powders that conserved a considerable proportion of anthocyanins with a process much more economical than with freeze-drying.

Although it is well known that the spectrophotometric method is not specific, in the present work we have demonstrated that this method allowed assessing differences between drying conditions or fruit species in a quicker,
less time-consuming and simple way than with HPLC-DAD (Pearson coefficient= 0.97, P<0.0001). This is important as
a first attempt for screening since identifying the anthocyanin compounds present or lost in each fruit or treatment
applied is still required to better understand results.

2.3.3.2.- Total polyphenol content

Statistical analysis based on the mixed model proposed showed a significant berry species-dependent source
strength change pattern of total polyphenol and antioxidant activity in berries for all drying conditions applied.
Total polyphenol content (TPC) of dried berries was analyzed in two solvent mixtures as described in the materials
and methods section. Table 6 shows the results obtained and can be seen that contrary to that observed for
anthocyanin content, the drying processes increased or decreased TPC, being dependent on the fruit species and
conditions applied.

Total polyphenol content (TPC) in acidified acetone extract (AcE) of dried fruits was higher than in acidified
methanolic extracts (MeE) in almost all samples (Table 6). These could be ascribed to the fact that MeE extracts were
processed to reduce sugar and interferences in HPLC-DAD analysis, which are known that react with Folin-Ciocalteu
reagent. Aqueous acetone has also been recognized as being more efficient in the extraction of condensed
polyphenols.

Accordingly, TPC in MeE for all dried berries in all conditions was higher than the corresponding freeze-dried
counterpart which could be explained by the reduction of interferences in these extracts, allowing the increase in
polyphenols due to release of bounded compounds (Table 6). These phenomena have also been reported in grape
pomace and seeds, cranberry pomace and hazel. Drying at 50 °C significantly decreased TPC in all fruits in AcE, in agreement with other studies reporting polyphenol
degradation during berry convective drying. Boysenberry maintained almost half the TPC, as compared to
freeze-dried samples, while the other berries lost around 70%.

At 65 °C TPC significantly increased in all berries species (P<0.05), particularly for Rubus genus in both extracts. This
drying condition with higher air temperature and shorter drying times than lyophilized or 50 °C resulted in an
increased content of compounds able to react with a Folin-Ciocalteu reagent, reducing substances and nitrogen-containing compounds formed during temperature-dependent processing. Hence, the increase observed in TPC
may probably be attributed to oxidation reactions of hydroxyl groups that produced more polyphenols or to
conversion of monophenols to di- or tri-phenols. As a result, the application of intermediate drying temperatures
such as 65 °C showed an almost complete conservation for *Ribes* or an increased polyphenol content for *Rubus*, indicating that phenolic compounds in berries varied in structure and stability associated with each genus selected. Berries dried at 130 °C showed that the increase in temperature was such that the degradation of polyphenols, interferences and Maillard products were also lost, resulting in an almost complete destruction of compounds capable of reacting with Folin-Ciocalteu reagent (4 to 17% of retention). These changes may affect the reactivity of aromatic rings, which could explain the decrease in the polyphenol content measured in the presence of Folin-Ciocalteu reagent in AcE from berries dried at 130 °C. It is remarkable that berries with blue color showed, in general, higher polyphenol content than the red ones in AcE it also holds true for MeE, with a few exceptions (Table 6). Differences found in TPC extracts reflect that polyphenols isolated during anthocyanin extraction by SPS solid extraction of the MeE included no significant amount of compounds that allow analyzing the effect of drying conditions, probably due to treatment to reduce interferences which eliminates most of Maillard products; thus the results were more related to berry species. As we have reported, the decrease in TPC observed at some drying conditions could be due to degradation of compounds, in which case a decrease in antioxidant activity is expected, even if it is well known that, for instance, Maillard reaction products have shown to have some antioxidant effect. On the other hand, the increase in TPC observed in fruits dried at 65 °C could be attributed to release of bounded polyphenols, other than anthocyanins, so that an increase or conservation of antioxidant potential is expected. As a result, the ABTS** radical cation scavenging activity and ferric-reducing power of AcE extracts were tested. We decided to use AcE extracts over MeE since that extraction was not purified, giving a more real measure of total antioxidant potential, even when it could be related, in part, to Maillard compounds, as reported by. 2.3.3.3.- Antioxidant activity determinations Many matrixes have been tested to study the effect of drying on antioxidant activity. Although the most important mechanism of antioxidant potential in berries is antiradical activity, in the present research the ferric-reducing power was also analyzed to further study the effect of different drying temperatures in both activities. Freeze-dried berries showed very similar ABTS** radical cation scavenging activity, purple fruits (boysenberry and blackcurrant) being the ones with the highest content (P<0.05) (Figure 6), although lyophilized fruits showed that blackcurrant was the berry with the highest reducing power, followed by boysenberry and redcurrant with very similar content between both, and raspberry with the lowest value. Convective drying of berries from *Ribes* genus
produced the almost complete loss of antioxidant activity considering both mechanisms: radical scavenging activity and reducing power with slight differences between them as previously described by other authors. In addition, ABTS$$^\ddagger$$ radical cation scavenging activity of Rubus fruits was maximum in all drying conditions tested. Convective drying at 50 °C produced slight loss of radical scavenging activity in Rubus genus, although this effect was not observed in the ferric-reducing power assay (Figure 6).

The increase of ABTS$$^\ddagger$$ radical cation scavenging activity in berries from Rubus genus was also observed for ferric reducing power in fruits dried at 65 °C. This observation agrees with the increase in polyphenol content observed after drying berry samples at 65 °C, which could be probably due to the depolymerization which increases free polyphenols. Additionally, polyphenols may be degraded during long-time drying (like the one performed at 50 °C) or by the use of high temperature (130 °C); as a result, both activities were highly affected (Figure 6), particularly in Ribes genus. This means that the best convective drying condition to produce dried Rubus berries has shown to be the one at 65 °C for 20 h, while no significant difference was found between drying Ribes fruits at 50 °C or 65 °C (except for BC).

ABTS$$^\ddagger$$ radical cation scavenging activity and ferric-reducing power showed significant correlation coefficients with total polyphenol content: 0.65 and 0.82, respectively (P<0.05). Additionally, significant correlation coefficients were observed between total anthocyanin and antiradical activity (0.40, P= 0.0012) and with ferric-reducing power (0.71, P<0.0001).

In conclusion, anthocyanin profiles of selected berries showed high predominance of delphinidin and cyanidin derivatives. Particularly, cyanidin 3-sophoroside derivatives and cyanidin 3-glucoside in Rubus genus and delphinidin 3-rutinoside and cyanidin-3-rutinoside in Ribes genus. As far as we know, there is scarce information available on the anthocyanin profile of these particular species, specifically from blackberry hydride, which constitutes a valuable contribution.

The study of the degradation of bioactive compounds in dried fruits is complex and dependent on the conditions used during drying. Better understanding of the best method of fruits drying for better preservation of bioactive compounds is required not only to avoid their loss during processing and storage of food, but also to determine possible implications to human health. In addition, the use of a simple and low-cost drying method such as a convective drying is a good alternative to include the dried product in other food matrixes without considerable increase of costs. Alterations in color, anthocyanin, total polyphenolic, radical scavenging activity and reducing power showed a major effect caused by drying conditions (time and temperature), and their levels were dependent...
on the particular red fruit. In particular, drying berries at 65 °C during 20 h was the best choice for the conservation of color, polyphenol content and antioxidant activity while anthocyanin content was similar in berries dried at 50 °C or 65 °C. It is clear that drying at temperature above 100 °C greatly deteriorates all the berry properties analyzed. It is remarkable that berries dried at 65 °C could be a great option to produce berry powder in order to improve the nutritional and sensorial characteristics of the final products. Conventional drying is a much more economical drying method than lyophilization and shows a considerable increase of total polyphenol content due to depolymerization of native polyphenols that also leads to increased antioxidant activity with pleasant smell and color.

REFERENCES


Figure 1.- Berries drying procedure.
Figure 2.- Analytical protocol for anthocyanin profiling, total polyphenol content and antioxidant activity determinations.
Figure 3.- Color parameters of dried berries at different conditions. Bubbles sized are related to redness (a*) value.

Raspberry (RB), boysenberry (BB), redcurrant (RC) and blackcurrant (BC).
Figure 4.- Chromatogram at 520nm corresponding to sample boysenberry (BB) lyophilized (left) and MSMS spectra of the peak eluting at 15.3 min with a M+ of 593.16 that has been identified as Cyanidin 3-(6''-dioxalyl-glucoside) (right). Cy 3-g, Cyanidin 3-glucoside; Cy 3-x, Cyanidin 3-xyloside; Cy 3-(6''-d-g), Cyanidin 3-(6''-dioxalyl-glucoside) and Cy 3-(6''-s-gl), Cyanidin 3-(6''-succinyl-glucoside).
Figure 5.- Total anthocyanin by HPLC-DAD (bars) and monomeric anthocyanin (dots) of freeze-dried berries (blue) and berries dried at 50 °C (yellow), 65 °C (red) and 130 °C (green).

Raspberry (RB), boysenberry (BB), redcurrant (RC) and blackcurrant (BC). *logarithmic scale
Figure 6.- ABTS•⁺ radical cation scavenging activity (bars) and ferric-reducing power (dots) of freeze-dried berries (blue) and berries dried at 50 °C (yellow), 65 °C (red) and 130 °C (green).

Raspberry (RB), boysenberry (BB), redcurrant (RC) and blackcurrant (BC).
<table>
<thead>
<tr>
<th></th>
<th>Water content</th>
<th>Ash</th>
<th>Protein</th>
<th>Carbohydrate</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>Raspberry</strong></td>
<td>85.8 ± 0.4c</td>
<td>1.68 ± 0.03a</td>
<td>4.7 ± 0.1d</td>
<td>7.9 ± 0.5a</td>
</tr>
<tr>
<td>USDA database*</td>
<td>85.75</td>
<td>-</td>
<td>1.20</td>
<td>11.94</td>
</tr>
<tr>
<td><strong>Boysenberry</strong></td>
<td>83.0 ± 0.4b</td>
<td>1.64 ± 0.02a</td>
<td>3.6 ± 0.0c</td>
<td>11.7 ± 0.4b</td>
</tr>
<tr>
<td>USDA database*</td>
<td>88.15</td>
<td>-</td>
<td>1.39</td>
<td>9.61</td>
</tr>
<tr>
<td><strong>Redcurrant</strong></td>
<td>82.4 ± 0.8b</td>
<td>2.06 ± 0.04b</td>
<td>2.8 ± 0.0b</td>
<td>12.8 ± 0.7b</td>
</tr>
<tr>
<td>USDA database*</td>
<td>83.95</td>
<td>-</td>
<td>1.40</td>
<td>13.80</td>
</tr>
<tr>
<td><strong>Blackcurrant</strong></td>
<td>75.4 ± 0.3a</td>
<td>2.93 ± 0.02c</td>
<td>3.1 ± 0.2b</td>
<td>18.6 ± 0.1c</td>
</tr>
<tr>
<td>USDA database*</td>
<td>81.96</td>
<td>-</td>
<td>1.40</td>
<td>15.38</td>
</tr>
</tbody>
</table>

*Within the same column, values with different letters indicate significant differences (P<0.05).*¹

USDA standard value for reference: https://ndb.nal.usda.gov/ndb/search/list
<table>
<thead>
<tr>
<th>Condition/Fruit</th>
<th>Raspberry</th>
<th>Boysenberry</th>
<th>Redcurrant</th>
<th>Blackberry</th>
</tr>
</thead>
<tbody>
<tr>
<td>Freeze-dried</td>
<td>0.376 ± 0.002&lt;sup&gt;d&lt;/sup&gt;</td>
<td>0.357 ± 0.001&lt;sup&gt;c&lt;/sup&gt;</td>
<td>0.242 ± 0.000&lt;sup&gt;a&lt;/sup&gt;</td>
<td>0.334 ± 0.001&lt;sup&gt;b&lt;/sup&gt;</td>
</tr>
<tr>
<td>50 °C</td>
<td>0.310 ± 0.002&lt;sup&gt;c&lt;/sup&gt;</td>
<td>0.413 ± 0.002&lt;sup&gt;d&lt;/sup&gt;</td>
<td>0.411 ± 0.002&lt;sup&gt;d&lt;/sup&gt;</td>
<td>0.398 ± 0.002&lt;sup&gt;d&lt;/sup&gt;</td>
</tr>
<tr>
<td>65 °C</td>
<td>0.251 ± 0.001&lt;sup&gt;a&lt;/sup&gt;</td>
<td>0.326 ± 0.001&lt;sup&gt;b&lt;/sup&gt;</td>
<td>0.342 ± 0.003&lt;sup&gt;c&lt;/sup&gt;</td>
<td>0.343 ± 0.002&lt;sup&gt;c&lt;/sup&gt;</td>
</tr>
<tr>
<td>130 °C</td>
<td>0.302 ± 0.001&lt;sup&gt;b&lt;/sup&gt;</td>
<td>0.280 ± 0.001&lt;sup&gt;a&lt;/sup&gt;</td>
<td>0.291 ± 0.001&lt;sup&gt;b&lt;/sup&gt;</td>
<td>0.282 ± 0.001&lt;sup&gt;a&lt;/sup&gt;</td>
</tr>
</tbody>
</table>
Table 3.- Chroma (ΔC<sub>ab</sub>) and total color difference (ΔE) for fruits dehydrated by convective drying*

<table>
<thead>
<tr>
<th>Fruit sample</th>
<th>Air-drying temperature</th>
<th>50 °C</th>
<th>65 °C</th>
<th>130 °C</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td>RB</td>
<td>BB</td>
<td>RC</td>
</tr>
<tr>
<td>ΔC&lt;sub&gt;ab&lt;/sub&gt;</td>
<td>24.3&lt;sup&gt;e&lt;/sup&gt;</td>
<td>10.7&lt;sup&gt;b&lt;/sup&gt;</td>
<td>31.6&lt;sup&gt;h&lt;/sup&gt;</td>
<td>13.0&lt;sup&gt;i&lt;/sup&gt;</td>
</tr>
<tr>
<td>ΔE</td>
<td>24.8&lt;sup&gt;e&lt;/sup&gt;</td>
<td>12.2&lt;sup&gt;b&lt;/sup&gt;</td>
<td>33.8&lt;sup&gt;e&lt;/sup&gt;</td>
<td>14.8&lt;sup&gt;c&lt;/sup&gt;</td>
</tr>
<tr>
<td></td>
<td>24.5&lt;sup&gt;b&lt;/sup&gt;</td>
<td>8.3&lt;sup&gt;a&lt;/sup&gt;</td>
<td>26.3&lt;sup&gt;f&lt;/sup&gt;</td>
<td>12.4&lt;sup&gt;c&lt;/sup&gt;</td>
</tr>
<tr>
<td>ΔC&lt;sub&gt;ab&lt;/sub&gt;</td>
<td>29.9&lt;sup&gt;e&lt;/sup&gt;</td>
<td>12.1&lt;sup&gt;c&lt;/sup&gt;</td>
<td>31.7&lt;sup&gt;h&lt;/sup&gt;</td>
<td>15.7&lt;sup&gt;d&lt;/sup&gt;</td>
</tr>
<tr>
<td>ΔE</td>
<td>34.1&lt;sup&gt;e&lt;/sup&gt;</td>
<td>14.5&lt;sup&gt;c&lt;/sup&gt;</td>
<td>33.1&lt;sup&gt;g&lt;/sup&gt;</td>
<td>17.0&lt;sup&gt;d&lt;/sup&gt;</td>
</tr>
</tbody>
</table>

*Within the same row, values with different letters indicate significant differences (P<0.05).

Raspberry (RB), boysenberry (BB), redcurrant (RC) and blackcurrant (BC).
Table 4.- Anthocyanin profile in lyophilized berries.

<table>
<thead>
<tr>
<th>Peak (n)</th>
<th>Rt (min)</th>
<th>Structure</th>
<th>M+ (Da)</th>
<th>Fragments (Da)</th>
<th>fruit</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>7.7</td>
<td>Cyanidin-3-sophoroside</td>
<td>611.16</td>
<td>449.09, 287.05</td>
<td>RB</td>
</tr>
<tr>
<td>2</td>
<td>7.7</td>
<td>Delphinidin 3-rutinoside</td>
<td>611.16</td>
<td>465.09, 303.04</td>
<td>BC</td>
</tr>
<tr>
<td>3</td>
<td>8.3</td>
<td>Cyanidin 3-sophoroside-5-rhamnoside</td>
<td>757.21</td>
<td>433.1, 287.05</td>
<td>RB</td>
</tr>
<tr>
<td>4</td>
<td>9.2</td>
<td>Cyanidin 3-glucoside</td>
<td>449.11</td>
<td>287.05</td>
<td>RB, BB</td>
</tr>
<tr>
<td>5</td>
<td>9.2</td>
<td>Cyanidin 3-xylosyl-rutinoside</td>
<td>727.2</td>
<td>287.06</td>
<td>RC</td>
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<tr>
<td>6</td>
<td>9.7</td>
<td>Cyanidin 3 -rutinoside</td>
<td>595.16</td>
<td>449.1, 287.05</td>
<td>RC, BC</td>
</tr>
<tr>
<td>7</td>
<td>12.8</td>
<td>Cyanidin 3-xyloside</td>
<td>419.05</td>
<td>287.05</td>
<td>BB</td>
</tr>
<tr>
<td>8</td>
<td>15.3</td>
<td>Cyanidin 3-(6''-dioxalyl-glucoside)</td>
<td>593.16</td>
<td>287.05</td>
<td>BB</td>
</tr>
<tr>
<td>9</td>
<td>18.4</td>
<td>Cyanidin 3-(6''-succinyl-glucoside)</td>
<td>549.12</td>
<td>287.05</td>
<td>BB</td>
</tr>
</tbody>
</table>

Raspberry (RB), boysenberry (BB), redcurrant (RC) and blackcurrant (BC).
Table 5.- Quantification of anthocyanins in dried fine fruits.

<table>
<thead>
<tr>
<th>Drying conditions</th>
<th>Fruit</th>
<th>Cy 3- sophoroside + Cy 3- sophoroside 5-rhamnoside</th>
<th>Cy 3- rutinoside</th>
<th>Cy 3- glucoside</th>
<th>D 3- rutinoside</th>
<th>Cy 3- xylosyl-rutinoside</th>
<th>Cy 3- xyloside</th>
<th>Cy 3- (6''-dioxalyl-glucoside)</th>
<th>Cy 3- (6''-succinyl-glucoside)</th>
</tr>
</thead>
<tbody>
<tr>
<td>FDB</td>
<td>RB</td>
<td>744±60</td>
<td>676±7*</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td></td>
<td>BB</td>
<td></td>
<td>3255±57</td>
<td></td>
<td>145±33</td>
<td>157±50</td>
<td>109±28</td>
<td></td>
<td></td>
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<tr>
<td></td>
<td>RC</td>
<td></td>
<td>427±33</td>
<td>1145±22</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td></td>
<td>BC</td>
<td>2512±282</td>
<td>2305±40</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>DB-50</td>
<td>RB</td>
<td>420±41</td>
<td>380±29*</td>
<td></td>
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<tr>
<td></td>
<td>BB</td>
<td></td>
<td>713±24</td>
<td>27.7±1</td>
<td>15.1±0</td>
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<td></td>
<td></td>
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<tr>
<td></td>
<td>RC</td>
<td></td>
<td>53±5</td>
<td>341±34</td>
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<td></td>
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<tr>
<td></td>
<td>BC</td>
<td>2338±64</td>
<td>2227±14</td>
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<td></td>
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<tr>
<td>DB-65</td>
<td>RB</td>
<td>187±14</td>
<td>148±52*</td>
<td></td>
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<td></td>
<td>BB</td>
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<td>882±12</td>
<td>1±0</td>
<td>65±0</td>
<td>23±0</td>
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<tr>
<td></td>
<td>RC</td>
<td></td>
<td>52±0</td>
<td>229±1</td>
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<tr>
<td></td>
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<tr>
<td>DB-130</td>
<td>RB</td>
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<td></td>
<td>BB</td>
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<td>37±12</td>
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<td>12±1</td>
<td>5±0</td>
<td>21±0</td>
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<tr>
<td></td>
<td>RC</td>
<td></td>
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<td></td>
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</tr>
<tr>
<td></td>
<td>BC</td>
<td>8±0</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td>6±0</td>
<td></td>
<td></td>
</tr>
</tbody>
</table>


Mean ± SD (n ≥ 3). Numbers in the table are in bold to show the most abundant anthocyanin in each sample, while white spaces correspond to not quantifiable anthocyanin. Within the same column, value with different letters indicate significant differences (P<0.05). Nd: not detectable.

*In RB Cy 3-glucoside co-eluted with Cy 3-rutinoside.
### Table 6.- Total polyphenol content in acetone and methanolic extracts of dried berries*

<table>
<thead>
<tr>
<th>Drying conditions</th>
<th>Fruit</th>
<th>Acidified acetone extract</th>
<th>Acidified methanolic extract</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>RB</td>
<td>18.7³</td>
<td>3.2³</td>
</tr>
<tr>
<td></td>
<td>BB</td>
<td>23.8³</td>
<td>5.6³</td>
</tr>
<tr>
<td></td>
<td>RC</td>
<td>21.5³</td>
<td>2.0³</td>
</tr>
<tr>
<td></td>
<td>BC</td>
<td>32.3³</td>
<td>3.9³</td>
</tr>
<tr>
<td>FDB</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td></td>
<td>RB</td>
<td>5.1³(-73%)</td>
<td>6.7³(+109%)</td>
</tr>
<tr>
<td></td>
<td>BB</td>
<td>12.5³(-47%)</td>
<td>6.6³(+18%)</td>
</tr>
<tr>
<td></td>
<td>RC</td>
<td>5.8³(-73%)</td>
<td>2.2³(+10%)</td>
</tr>
<tr>
<td></td>
<td>BC</td>
<td>11.6³(-64%)</td>
<td>11.0³(+182%)</td>
</tr>
<tr>
<td>DB50</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td></td>
<td>RB</td>
<td>35.2³(+88%)</td>
<td>5.4³(+69%)</td>
</tr>
<tr>
<td></td>
<td>BB</td>
<td>41.6³(+75%)</td>
<td>12.0³(+114%)</td>
</tr>
<tr>
<td></td>
<td>RC</td>
<td>23.1³(+7%)</td>
<td>3.8³(+90%)</td>
</tr>
<tr>
<td></td>
<td>BC</td>
<td>37.2³(+15%)</td>
<td>7.7³(+97%)</td>
</tr>
<tr>
<td>DB65</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td></td>
<td>RB</td>
<td>1.4³(-93%)</td>
<td>6.7³(+109%)</td>
</tr>
<tr>
<td></td>
<td>BB</td>
<td>4.0³(-83%)</td>
<td>10.9³(+95%)</td>
</tr>
<tr>
<td></td>
<td>RC</td>
<td>3.6³(-83%)</td>
<td>3.9³(+95%)</td>
</tr>
<tr>
<td></td>
<td>BC</td>
<td>5.8³(-82%)</td>
<td>4.3³(+10%)</td>
</tr>
</tbody>
</table>


*Within the same column, values with different letters indicate significant differences (P<0.05)

Values between parentheses indicate percentage of increase or decrease with respect to FDB.