

## The influence of different air-drying conditions on bioactive compounds and antioxidant activity of berries

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**13 ABSTRACT**

14 The aim of the present research was to study the effect of convective drying on color, bioactive compounds and  
15 antioxidant activity of berry fruits and to chemically characterize the polyphenolic composition of raspberry,  
16 boysenberry, redcurrants and blackcurrants fruit. Drying berries at 65 °C provoked the best conservations of color,  
17 particularly for boysenberry and blackcurrant. Drying at 65 °C was also the condition that showed higher level of  
18 polyphenols, while drying at 50 °C or 130 °C showed above % degradation of them due to the long time or high  
19 temperature drying. Radical scavenging activity was the predominant antioxidant mechanism in all samples, being 65  
20 °C dried berries the most active ones possibly due to polyphenols depolymerization. The anthocyanin profile showed  
21 that delphinidin and cyanidin derivatives were the most abundant anthocyanidins with different predominance  
22 between berry genera. Degradation of anthocyanins was increased with drying temperature been Cy 3-glucoside and  
23 Cy 3-rutinoside the most abundant.

**24 1.- INTRODUCTION**

25 Berry fruits are a type of small fine fruits characterized by a red, purple and blue color. The most common are:  
26 blueberry, cranberry, blackberry, raspberry, white, red or blackcurrant and strawberry. The consumption of these  
27 fruits is mainly restricted to fresh product or processed food such as juice, beverage and jam due to their short shelf-  
28 life, so that different marketing strategies like drying and/or packaging can be used to retain the quality of berries  
29 during storage <sup>1</sup>. In this sense, dried or frozen berries are the two most commonly consumed forms of these fruits  
30 despite the decrease in nutritional value found in the former or the high cost of transportation in the latter.

31 Berries contain high levels of polyphenols including flavonoids (anthocyanins, flavonols and flavanols such as  
32 condensed tannins or proanthocyanidins), hydrolysable tannins (ellagitannins and gallotannins), phenolic acids  
33 (hydroxybenzoic and hydroxycinnamic acids, chlorogenic acid), stilbenoids and lignans <sup>2,3</sup>. Many studies have been  
34 published in relation to the health benefits of berry consumption due to the presence of those active compounds <sup>4-6</sup>.  
35 Thanks to the rich and diverse composition of bioactive compounds and their health-promoting properties which  
36 result mostly from their antioxidant activity, their metal chelating capacity and their affinity for proteins, berry fruits  
37 are widely recognized as natural functional products <sup>7,8</sup>.

38 Since the availability of berries is seasonal, their processing for long-term storage and their addition to the  
39 formulation of new products are a good way to improve the nutritional quality of final products. The most commonly  
40 used methods of food preservation involve thermal treatments to reduce moisture content to a safe level, depriving

41 molds of favorable proliferative conditions <sup>9,10</sup>. In this sense, convective drying is always an alternative to extend  
42 shelf life and allow powders to be used as additives of other food matrixes, promoting the use of these types of  
43 highly nutritious fruits <sup>11,12</sup>.

44 It should be noted that quality characteristics of dehydrated fruits will be affected by drying conditions, in which  
45 phytochemicals like anthocyanins and phenolic compounds are highly susceptible to degradation <sup>13</sup>, thus reducing  
46 their antioxidant activity, and in general, their functional characteristics. Some studies related to drying berries  
47 indicate that this technological process may have negative effects on bioactive compounds compared to their fresh  
48 counterpart <sup>11</sup>. Therefore, the conditions of the drying process can produce light or moderate effects on fruit's  
49 bioactive compounds, mainly depending on the type of drying process and the fruit species <sup>14</sup>. As a result, dried fruits  
50 could not only be a good source of vitamins and fiber but also provide a wide array of bioactive phytochemicals that  
51 have been linked to a reduction in the risk of chronic diseases. Profiling the bioactive compounds that remains in  
52 dried fruits is the first step to address benefits associated to their consumption.

53 Since drying process affects appearance and chemical composition, the best method should be selected to ensure  
54 maximum conservation of bioactive compounds. Freeze drying is probably the best technique to reach high quality  
55 dried fruits; however, it is also one of the most expensive ones <sup>15</sup>. Moreover, freeze drying is the best method for  
56 obtaining a high-quality dried product (for heat sensitive materials) since it is commonly considered that minor  
57 modifications take place <sup>16</sup>. On the other hand, convective drying is the most economical and widely adopted  
58 technique in the food industry, despite requiring long drying times and high temperatures <sup>11</sup>. In the last years, new  
59 emerging technologies for drying fruits have been studied <sup>9</sup>, although due to fine fruits like berries are commonly  
60 more expensive than others, their applications increases the cost even more. This fact reinforces the study of  
61 traditional drying technologies in order to determine the best conditions to preserve the characteristics of dried  
62 fruits that could be used to food enrichment without considerable costs increase of final product. Additionally, the  
63 effects of drying on polyphenolics and antioxidant activities have not been systematically studied, this selecting the  
64 best conditions to maximize presentation of color, anthocyanins and antioxidant activity, is crucial to produce dried  
65 fruit powder to be used as food coloring, snack production or functional ingredients to be included in other food  
66 products. Therefore, the aim of this work was to study the impact of convective drying at different temperatures  
67 those characterizes of berry fruits and to chemically characterize their polyphenolic composition.

## 68 **2.- MATERIALS AND METHODS**

### 69 **2.1.- Materials**

70 The berry fruits for the present research were selected from four species, each two belonging to a different genus.  
71 Raspberry (*Rubus idaeus* var. Autumn Bliss) and boysenberry (*R. ursinus* × *R. idaeus* var. Black Satin) were chosen  
72 from *Rubus* genus, and redcurrants (*Ribes rubrum* sp.) and blackcurrants (*Ribes nigrum* sp.) from *Ribes* genus. All  
73 berries were purchased from Dolphes Gourmet (Rosario, Argentina) as Individual Quick Frozen (IQF) fruits, assuring  
74 low-quality changes and longer conservation times. *Rubus* genus fruits came from San Pedro, Buenos Aires  
75 (Argentina) while *Ribes* genus fruits from El Bolsón, Río Negro (Argentina). Fruits from two different crop years were  
76 purchased and combined in two pools considered as duplicates (raspberry and boysenberry from 2014-2015 and  
77 redcurrants and blackcurrants from 2013-2014).

78 All chemicals were of analytical grade unless otherwise stated. Formic and chlorhidric acids, methanol and acetonitrile  
79 (HPLC grade) were obtained from Fisher Scientific (Madrid, Spain). Delphinidin-3-glucoside, cyanidin-3-glucoside,  
80 peonidin-3-glucoside, pelargonidin-3-glucoside and malvidin-3-glucoside standards were purchased from  
81 Extrasynthese (Lyon, France).

## 82 **2.2.- Methods**

### 83 2.2.1.- Physicochemical characteristics of selected berry fruits

84 Water content (method 934.06), ash (method 930.35) and proteins (method 920.152) were determined according to  
85 AOAC methods<sup>17</sup>. It should be noted that fruits and their seeds were analyzed.

### 86 2.2.2.- Berries drying

87 Figure 1 shows the procedure used for drying berries. Briefly, each berry sample was separated in three sets in order  
88 to apply different convective drying conditions: 50 °C for 48 h, 65 °C for 20 h or 130 °C for 2 h until a moisture  
89 content below 15% was obtained. Until that, berries were blanched for 3 min in a home steamer (Smart-Tek SD2071,  
90 Argentina) for better conservation of anthocyanins and polyphenols and inactivation of polyphenol oxidase from  
91 berries as reported by Sablani *et al.*,<sup>15</sup>. The blanched fruits were carefully placed in a plastic mesh inside the air  
92 convection drier and separated in three portions and were exposed to the previously described conditions.  
93 Additionally, freeze-dried berries were obtained by storage at -80 °C and freeze dried for 48 h (L-T8 RIFICOR,  
94 Argentina). These samples were used as a reference assuming no significant modifications induced by lyophilization  
95 method. All samples were ground in a coffee mill (PE-MC9100, Peabody, Argentina) and stored at -18 °C prior to  
96 analysis. Water activity was measured by using an electric hygrometer Novasina Lab MASTER-aw (Novasina AG,

97 Lanchen, Switzerland) at 25°C. Measurements were done in duplicate and are expressed as the media +- standard  
98 deviation.

### 99 2.2.3.- Color of dried berries

100 The color of dried fruit powders was measured with a spectrophotometer (Minolta, Ramsey, NJ). Eight-millimeter  
101 measurement apertures, D65 illuminant, 10° angle of observer was settled and color recorded in CIE Lab space. L\*  
102 indicates lightness, its value ranging from 0 (black) to 100 (white); a\* and b\* are the chromaticity coordinates. From  
103 the CIELAB coordinates, color function chroma ( $\Delta C_{ab}$ ) and total color change ( $\Delta E$ ) were calculated according to the  
104 following equations:

$$105 \quad \Delta C_{ab} = (\Delta a^{*2} + \Delta b^{*2})^{0.5} \quad \text{eq. 1}$$

$$106 \quad \Delta E = (\Delta L^{*2} + \Delta a^{*2} + \Delta b^{*2})^{0.5} \quad \text{eq. 2}$$

### 107 2.2.4.- Bioactive compounds and antioxidant activity in dried berries

#### 108 2.2.4.1.- Anthocyanin analysis

##### 109 2.2.4.1.1.- Anthocyanin profile by HPLC-DAD-ESI/MS-QTOF

110 The analytical protocol for anthocyanins profile determination of the different samples studied is presented in Figure  
111 2. Briefly, the method of anthocyanin purification was performed according to García-Herrera *et al.*<sup>18</sup>. A hydro-  
112 alcoholic extraction was done and samples were purified by an aqueous extraction followed by a amethanolic one,  
113 both extracts were mixed and passed though Agilent Tech cartridge (Figure 2). This extract was used to obtain the  
114 anthocyanin profile by HPLC analysis carried out on a liquid chromatography system (Hewlett Packard Agilent 1200  
115 Series) equipped with a quaternary pump and a photo- diode array detector (DAD) (Agilent Technologies). The  
116 column used was a Phenomenex Luna C18 column (5  $\mu\text{m}$ , 4.6 mm x 150 mm), set thermostatically at 25 °C.  
117 Chromatographic data were acquired and processed using an Agilent Chemstation for LC 3D system (Rev. B.04.01,  
118 Agilent Technologies). Briefly, the binary mobile phase used for analysis was aqueous 0.1% formic acid and HPLC-  
119 grade acetonitrile at a flow rate of 0.5 mL min<sup>-1</sup>. Samples were analyzed in triplicate. Peaks were identified by  
120 comparing their retention time (Rt) and UV- visible spectra with the reference compounds, and the data were  
121 quantified using the corresponding curves of the reference compounds as standards, when available.

122 To confirm the identity of the compounds recorded, additional analyses were performed using HPLC coupled with  
123 mass spectrometry detection (HPLC-MS- QTOF): liquid chromatography/mass selective Agilent 1200: quaternary  
124 pump (G1311A), diode array detector (G1315B). The column used was an Phenomenex Luna C18 column (5  $\mu\text{m}$ , 4.6  
125 mm x 150 mm); Mass QTOF (SAcetopetrometre Agilent G6530A Accurate Mass Q- TOF LCMS) with Electrospray  
ACS Paragon Plus Environment

126 Ionization (ESI) with JetStream technology; Instrument State: standard dynamic range m/z 3200. Software used was  
127 Masshunter Data Acquisition B.04.00 and Masshunter Qualitative Analysis B.04.00.

#### 128 2.2.4.1.2. Monomeric anthocyanin content

129 Four different solvent mixtures were prepared in order to extract the largest quantity of bioactive compounds from  
130 the whole fruits. Fifty milligrams of freeze-dried berries or 500 mg of air-dried were mixed with 1.5 ml of each  
131 solvent mixture: methanol: water (70:30), methanol: water (50:50), ethanol: water (70:30) and acetone: water  
132 (70:30) in all cases with a final concentration of 0.1% HCl. The solvent/berry mixtures were mixed for 10 min and  
133 then centrifuged at 12,000 g for 15 min. The supernatant was recovered and seven more extractions were  
134 performed. All determinations were made in the supernatants recovered. The solvent with better extraction  
135 performance based on the total anthocyanin content method was acetone: water (70:30) (data not shown), thus  
136 selected to determine total polyphenol content and antioxidant activity analysis (Figure 2).

137 Total anthocyanin content was performed according to Giusti and Wrolstad (2001) in the acidified dried berry  
138 extracts using an extinction coefficient (B) of  $26,900 \text{ l cm}^{-1} \text{ mg}^{-1}$  and a molecular weight (MW) of  $449.2 \text{ g/mol}$  of  
139 cyanidin 3-glucoside. Absorbance were measured at 520 nm ( $A^{520}$ ) and 700 nm ( $A^{700}$ ) and calculated as follows:

$$140 \quad \text{Cyanidin 3-glucoside } \mu\text{g/g dried fruit} = [(A^{520} - A^{700})_{\text{pH } 1} - (A^{520} - A^{700})_{\text{pH } 4.5}] \times \text{MW} \times F \times V_e \times 10^5 / (B \times 1 \times W_s),$$

141 F being the dilution factor for sample,  $V_e$  the extract volume and  $W_s$  the sample weight.

#### 142 2.2.4.2.- Total polyphenol content

143 Total polyphenol content of dried berries was determined in two extracts as indicated in Figure 2, that was the  
144 methanolic extract made for anthocyanin profiling ) and in the acidified acetone extract for extraction of bioactive  
145 components previously described. Total polyphenols were determined using the Folin-Ciocalteu method, with gallic  
146 acid as a calibration standard <sup>20</sup>. The concentration of total polyphenols was expressed as mg gallic acid per gram of  
147 fruit powder.

#### 148 2.2.4.3.- Antioxidant activity determinations

##### 149 2.2.4.3.1.- ABTS<sup>•+</sup> radical cation scavenging activity

150 The ABTS<sup>•+</sup> radical cation scavenging activity was measured according to Re *et al.*<sup>21</sup> . Trolox (Sigma 238813) was used  
151 as a standard and results were expressed as  $\mu\text{mol}$  of Trolox equivalent per gram of fruit powder.

##### 152 2.2.4.2.3.- Ferric-reducing ability

153 Ferric reducing activity of dried fruits was determined by FRAP assay according to Pulido *et al.*,<sup>22</sup> using gallic acid as a  
154 standard.

### 155 2.2.5.- Statistical analysis

156 All samples were prepared in duplicate and each replicate was quantified in duplicate. Results were analyzed by the  
157 adjustment to a model with fixed effects for a classification factor with eight levels (drying methods and fruit type).  
158 The model included a variance function to take account of the presence of an increasing variability pattern related to  
159 medium levels of response variable. The adjustment was carried out using an implementation in InfoStat Software <sup>23</sup>  
160 of gls function from the nlme library <sup>24</sup> of R <sup>25</sup>. The variance function applied was a function of implementation of  
161 power variance varPower() from nlme library. This type of statistical analysis allows comparing, simultaneously, the  
162 effect of drying method and fruit type. Results of the analyses were evaluated by using DGC test <sup>26</sup> with a degree of  
163 significance of  $P < 0.05$ . Pearson correlation coefficients were used to determine the relationship between 166  
164 anthocyanins contents determined by chromatographic and spectrophotometric methods.

## 165 **2.3.- RESULTS AND DISCUSSION**

### 166 2.3.1.- Physicochemical characteristics of selected berry fruits

167 The selected berry fruits were characterized according to their physicochemical composition as shown in Table 1.  
168 The four berry fruits showed high water content, raspberry being the one with the highest value ( $P < 0.05$ ).  
169 Blackcurrant presented the highest ash content, almost twice that of the observed for raspberry and boysenberry.  
170 Protein content was significantly higher for raspberry compared with both currant and boysenberry. Carbohydrate  
171 content was twice higher for blackcurrant compared with raspberry, while boysenberry and redcurrant showed  
172 intermediate values. USDA database <sup>27</sup> was used as a reference for water content, protein and carbohydrate content  
173 of berries. Water content of selected berry fruits was very similar to the corresponding USDA value; yet, selected  
174 species showed increased protein content and slight difference in carbohydrate percentage compared to the  
175 standard value.

176 Water activity is a factor that highly influence dried fruit stability. A high water activity can lead to a shorter storage  
177 time of products, which is due to the possibility of microbial growth and biochemical changes, has been suggested a  
178 value bellow 0.600 to eliminate these factors <sup>28</sup>. In this sense, the water activity of dried berries at different  
179 conditions ranged from 0.242 to 0.413 (Table 2). Considering each condition applied for drying fruits the final water  
180 activity decreased with increasing drying temperature, showing the following order: freeze-dried (0.3269) > 50 °C  
181 (0.3828) > 65 °C (0.3156) > 130 °C (0.2883). Water absorption values was very similar between all dried fruit within  
182 one specie, being on average: 0.309 for raspberry, 0.344 for boysenberry, 0.321 for redcurrant and 0.339 for



183 blackcurrant, similar values was found by Samoticha *et al.*,<sup>29</sup> after convective drying of chokeberries. As a result, it  
184 can be assumed that the obtained dried berries powders were microbiologically stable.

185

### 186 2.3.2.- Color of dried berries

187 Color and appearance in berries attract the consumer, they being associated with higher hedonic responses in fruits  
188 with darker color or higher anthocyanin content<sup>1,30</sup>. Additionally, changes in color during thermal processing of fruit  
189 might provide information about alterations in the content of anthocyanins and other polyphenols. Statistical  
190 analysis showed that not only fruit type determined color variables but also drying condition influences them with a  
191 significant interaction in luminosity (L\*), redness (a\*) and yellowness (b\*).

192 Luminosity of dried fruits decreased with increasing drying temperature in all fruits, raspberry samples being those  
193 with highest L\* values in all cases. In addition, drying blackcurrants at all convective drying conditions generated a  
194 large decrease in L\* with no significant difference ( $p>0.05$ ) between convective treatments (Figure 3). As seen in  
195 Figure 3, in addition to the significant interaction between fruit species and drying conditions, freeze drying was the  
196 best option to obtain dried fruits with high luminosity (L\*= 34.46 on average), followed by drying at 65 °C (L\*= 29.52  
197 on average). On the other hand, convective drying at 130 °C decreased largely luminosity with a mean value of 24.31.  
198 From Figure 3 it is clear that redness was the most affected color parameter during convective drying of berries. In  
199 addition, slight differences were detected between fruit drying at 50°C or 65°C (a\*= 7.38 on average) compared to  
200 freeze-drying (a\*= 25.62 on average) and high temperature drying (a\*= 3.89 on average). That means that the drying  
201 method showed a main effect on redness compared to fruit species. As observed with L\* values, the highest redness  
202 was observed in freeze-dried berries. This could indicate that discoloration and browning during air drying may be  
203 the result of various chemical reactions including pigment destruction<sup>31</sup>.

204 Since selected fruits are red/purple, yellowness was the less affected color variable; only drying at 130 °C showed a  
205 clear effect with the lowest b\* values (b\*= 1.81 on average). Raspberry presented the highest yellowness in all  
206 drying conditions (except for freeze-dried conditions), while blackcurrant showed the minimum yellowness in all  
207 conditions. For *Rubus* genus drying at 50 °C and 65 °C increased yellowness compared to freeze-dried conditions  
208 (Figure 3).

209 For better analysis of color modifications due to fruit species or drying conditions, chroma ( $\Delta C_{ab}$ ) and total color  
210 difference ( $\Delta E$ ) were determined (Table 3) using freeze-drying condition as a reference. Since chroma included  
211 variations in redness and yellowness, while total color difference also included luminosity, the similar values

212 observed in both parameters indicated that L\* showed no considerable disparity between different dehydrated  
213 fruits species. Boysenberry showed the minimum chrome and total color differences in all conditions followed by  
214 blackcurrant, indicating that blue fruits better conserved color parameters, while major changes were found for  
215 raspberry dried at 130 °C.

### 216 2.3.3.- Bioactive compounds and antioxidant activity in dried berries

#### 217 2.3.3.1.- Anthocyanin analysis

218 Anthocyanin quantification was carried out using two methods: HPLC-DAD-ESI/MS-QTOF (anthocyanin profile) and a  
219 spectrophotometric estimation (monomeric anthocyanins expressed as µg cyanidin 3-glucoside). Anthocyanin profile  
220 allows identifying different compounds in each fruit species as shown in Table 4, which is particularly new for  
221 boysenberry (blackberry hydride) selected in presented research and shown in Figure 4. In this sense, three different  
222 anthocyanin compounds were found in raspberry while four were detected in boysenberry, cyanidin 3-glucoside  
223 being found in both fruits. Analysis of redcurrant (RC) and blackcurrant (BC) (*Ribes* genus) showed only two  
224 compounds found in each one, being cyaniding 3-rutinoside the one in common.

225 Quantification of each identified anthocyanin was performed and results shown in Table 5. Analysis of anthocyanin  
226 profile showed that freeze-drying was the condition with the highest anthocyanin content in all samples. On the  
227 other hand, air-drying at 130 °C destroys anthocyanins, quantification in both purple fruits (boysenberry and  
228 blackcurrant) being minimum, in agreement with the low values in total color difference and chroma results  
229 described above.

230 Cy-3-sophoroside was co-eluted with Cy-3-sophoroside-5-rhamnoside in our conditions and thus quantified  
231 together. These two cyanidin compounds were the major anthocyanins in raspberry, reaching 52.3% of total  
232 anthocyanin in lyophilized form, in agreement with other authors<sup>32,18,33</sup>. Compared with freeze-drying, only 56% and  
233 25% of those anthocyanins were conserved when drying at 50 °C and 65 °C was applied, respectively, while no  
234 anthocyanin was detected in raspberry dehydrated at 130 °C. A similar proportion of Cy-3-glucoside conservation at  
235 intermediate drying temperatures was also detected for that fruit.

236 The anthocyanin profile of boysenberry was consistent with those of previously reports in blackberries<sup>33-35</sup> with a  
237 single major peak representing Cy-3-glucoside that constituted 88.8% of the total anthocyanin content in the freeze-  
238 dried form. It should be noted that, in addition to decreasing anthocyanin content compared to freeze-drying  
239 conditions, dehydrating boysenberry by air-drying showed minor differences between 50°C and 65°C with 22% and

240 27% of Cy-3-glucoside conserved, respectively. In addition, in samples dried at 130°C, a minimum quantity of four  
241 anthocyanins was found.

242 Delphinin-3-rutinoside and Cy-3-xylosyl-rutinoside were present in redcurrant at all drying conditions (except for  
243 drying at 130°C), Cy-3-xylosyl-rutinoside accounting for around 72.8% of total anthocyanins. As observed for  
244 boysenberry, drying at 50°C and 65°C produced the similar loss of compounds compared to freeze-dried redcurrant  
245 (Table 5). Blackcurrant showed equal quantity of rutinoside derivatives conserved after drying at 50°C compared to  
246 lyophilized form.

247 Although no anthocyanin was found in blackcurrant dehydrated at 65 °C and a depreciable amount was found after  
248 drying at higher temperature, the profile observed in both *Ribes* genera was similar to those of other authors that  
249 reported delphinine-3-rutinoside in both fruits with higher content in blackcurrant than in redcurrant<sup>32,33,36,37</sup>.

250 Figure 5 shows that results similar to the total anthocyanin detected by HPLC-DAD method were obtained when  
251 estimating the monomeric anthocyanin concentration via the spectrophotometric method with a Pearson's  
252 correlation coefficient of 0.76 (P<0.05). In general, blackcurrant was the fruit with the highest anthocyanin content  
253 in all conditions (except drying at 65°C by HPLC-DAD method) followed by boysenberry, which could be attributed to  
254 the dark color of these berry species. Similar results were found by Arancibia-Avila, et al.<sup>13</sup> by comparing raspberry  
255 and blueberries, indicating that color differences were closely associated to anthocyanins content. In this regard,  
256 raspberry and redcurrant showed the lowest anthocyanin content, showing similar values between them. With both  
257 drying methods a detrimental effect of drying conditions was observed, from which freeze-drying was found to be  
258 the method which allowed conservation of the highest concentration of anthocyanins. Drying berries at 50 °C or 65  
259 °C produced fruits with similar anthocyanin content and the differences detected in freeze-dried berries according to  
260 each fruit was conserved but less pronounced. On the other hand, drying at 130 °C showed almost complete loss of  
261 anthocyanins (Figure 5).

262 In general, anthocyanin degradation seems to be more related to drying temperature than with exposure time since  
263 with increase in temperature, the time needed to dry fruits was reduced. As a result, anthocyanins were destroyed  
264 with increased drying temperature in a different proportion according to fruit species (significant interaction), in  
265 agreement with Karam *et al*,<sup>11</sup>. Drying berries at intermediate temperatures produced fruit powders that conserved  
266 a considerable proportion of anthocyanins with a process much more economical than with freeze-drying.

267 Although it is well known that the spectrophotometric method is not specific, in the present work we have  
268 demonstrated that this method allowed assessing differences between drying conditions or fruit species in a quicker,

269 less time-consuming and simple way than with HPLC-DAD (Pearson coefficient= 0.97,  $P < 0.0001$ ). This is important as  
270 a first attempt for screening since identifying the anthocyanin compounds present or lost in each fruit or treatment  
271 applied is still required to better understand results.

### 272 **2.3.3.2.- Total polyphenol content**

273 Statistical analysis based on the mixed model proposed showed a significant berry species-dependent source  
274 strength change pattern of total polyphenol and antioxidant activity in berries for all drying conditions applied.

275 Total polyphenol content (TPC) of dried berries was analyzed in two solvent mixtures as described in the materials  
276 and methods section. Table 6 shows the results obtained and can be seen that contrary to that observed for  
277 anthocyanin content, the drying processes increased or decreased TPC, being dependent on the fruit species and  
278 conditions applied.

279 Total polyphenol content (TPC) in acidified acetone extract (AcE) of dried fruits was higher than in acidified  
280 methanolic extracts (MeE) in almost all samples (Table 6). These could be ascribed to the fact that MeE extracts were  
281 processed to reduce sugar and interferences in HPLC-DAD analysis, which are known that react with Folin-Ciocalteu  
282 reagent <sup>38</sup>; Aqueous acetone has also been recognized as being more efficient in the extraction of condensed  
283 polyphenols <sup>39</sup>.

284 Accordingly, TPC in MeE for all dried berries in all conditions was higher than the corresponding freeze-dried  
285 counterpart which could be explained by the reduction of interferences in these extracts, allowing the increase in  
286 polyphenols due to release of bounded compounds (Table 6). These phenomena have also been reported in grape  
287 pomace and seeds <sup>40</sup>, cranberry pomace <sup>41</sup> and hazel <sup>42</sup>.

288 Drying at 50 °C significantly decreased TPC in all fruits in AcE, in agreement with other studies reporting polyphenol  
289 degradation during berry convective drying <sup>13,15</sup>. Boysenberry maintained almost half the TPC, as compared to  
290 freeze-dried samples, while the other berries lost around 70%.

291 At 65 °C TPC significantly increased in all berries species ( $P < 0.05$ ), particularly for *Rubus* genus in both extracts. This  
292 drying condition with higher air temperature and shorter drying times than lyophilized or 50 °C resulted in an  
293 increased content of compounds able to react with a Folin-Ciocalteu reagent, reducing substances and nitrogen-  
294 containing compounds formed during temperature-dependent processing <sup>38,43</sup>. Hence, the increase observed in TPC  
295 may probably be attributed to oxidation reactions of hydroxyl groups that produced more polyphenols or to  
296 conversion of monophenols to di- or tri-phenols <sup>12</sup>. As a result, the application of intermediate drying temperatures

297 such as 65 °C showed an almost complete conservation for *Ribes* or an increased polyphenol content for *Rubus*,  
298 indicating that phenolic compounds in berries varied in structure and stability associated with each genus selected.  
299 Berries dried at 130 °C showed that the increase in temperature was such that the degradation of polyphenols,  
300 interferences and Maillard products were also lost, resulting in an almost complete destruction of compounds  
301 capable of reacting with Folin-Ciocalteu reagent (4 to 17% of retention). These changes may affect the reactivity of  
302 aromatic rings, which could explain the decrease in the polyphenol content measured in the presence of Folin-  
303 Ciocalteu reagent in AcE from berries dried at 130 °C.

304 It is remarkable that berries with blue color showed, in general, higher polyphenol content than the red ones in AcE  
305 it also holds true for MeE, with a few exceptions (Table 6).

306 Differences found in TPC extracts reflect that polyphenols isolated during anthocyanin extraction by SPS solid  
307 extraction of the MeE included no significant amount of compounds that allow analyzing the effect of drying  
308 conditions, probably due to treatment to reduce interferences which eliminates most of Maillard products; thus the  
309 results were more related to berry species.

310 As we have reported, the decrease in TPC observed at some drying conditions could be due to degradation of  
311 compounds, in which case a decrease in antioxidant activity is expected, even if it is well known that, for instance,  
312 Maillard reaction products have shown to have some antioxidant effect<sup>44</sup>. On the other hand, the increase in TPC  
313 observed in fruits dried at 65 °C could be attributed to release of bounded polyphenols, other than anthocyanins, so  
314 that an increase or conservation of antioxidant potential is expected. As a result, the ABTS<sup>••</sup> radical cation  
315 scavenging activity and ferric-reducing power of AcE extracts were tested. We decided to use AcE extracts over MeE  
316 since that extraction was not purified, giving a more real measure of total antioxidant potential, even when it could  
317 be related, in part, to Maillard compounds, as reported by<sup>14</sup>.

### 318 **2.3.3.3.- Antioxidant activity determinations**

319 Many matrixes have been tested to study the effect of drying on antioxidant activity<sup>13,15,45</sup>. Although the most  
320 important mechanism of antioxidant potential in berries is antiradical activity, in the present research the ferric-  
321 reducing power was also analyzed to further study the effect of different drying temperatures in both activities.

322 Freeze-dried berries showed very similar ABTS<sup>••</sup> radical cation scavenging activity, purple fruits (boysenberry and  
323 blackcurrant) being the ones with the highest content (P<0.05) (Figure 6), although lyophilized fruits showed that  
324 blackcurrant was the berry with the highest reducing power, followed by boysenberry and redcurrant with very  
325 similar content between both, and raspberry with the lowest value. Convective drying of berries from *Ribes* genus

326 produced the almost complete loss of antioxidant activity considering both mechanisms: radical scavenging activity  
327 and reducing power with slight differences between them as previously described by other authors<sup>14,15,31</sup>. In  
328 addition, ABTS<sup>•+</sup> radical cation scavenging activity of *Rubus* fruits was maximum in all drying conditions tested.  
329 Convective drying at 50 °C produced slight loss of radical scavenging activity in *Rubus* genus, although this effect was  
330 not observed in the ferric-reducing power assay (Figure 6).

331 The increase of ABTS<sup>•+</sup> radical cation scavenging activity in berries from *Rubus* genus was also observed for ferric  
332 reducing power in fruits dried at 65 °C. This observation agrees with the increase in polyphenol content observed  
333 after drying berry samples at 65 °C, which could be probably due to the depolymerization which increases free  
334 polyphenols. Additionally, polyphenols may be degraded during long-time drying (like the one performed at 50 °C) or  
335 by the use of high temperature (130 °C); as a result, both activities were highly affected (Figure 6), particularly in  
336 *Ribes* genus. This means that the best convective drying condition to produce dried *Rubus* berries has shown to be  
337 the one at 65 °C for 20 h, while no significant difference was found between drying *Ribes* fruits at 50 °C or 65 °C  
338 (except for BC).

339 ABTS<sup>•+</sup> radical cation scavenging activity and ferric-reducing power showed significant correlation coefficients with  
340 total polyphenol content: 0.65 and 0.82, respectively ( $P < 0.05$ ). Additionally, significant correlation coefficients were  
341 observed between total anthocyanin and antiradical activity (0.40,  $P = 0.0012$ ) and with ferric-reducing power (0.71,  
342  $P < 0.0001$ ).

343 In conclusion, anthocyanin profiles of selected berries showed high predominance of delphinidin and cyanidin  
344 derivatives. Particularly, cyanidin 3-sophoroside derivatives and cyanidin 3-glucoside in *Rubus* genus and delphinidin  
345 3-rutinoside and cyanidin-3-rutinoside in *Ribes* genus. As far as we know, there is scarce information available on the  
346 anthocyanin profile of these particular species, specifically from blackberry hydride, which constitutes a valuable  
347 contribution.

348 The study of the degradation of bioactive compounds in dried fruits is complex and dependent on the conditions  
349 used during drying. Better understanding of the best method of fruits drying for better preservation of bioactive  
350 compounds is required not only to avoid their loss during processing and storage of food, but also to determine  
351 possible implications to human health. In addition, the use of a simple and low-cost drying method such as a  
352 convective drying is a good alternative to include the dried product in other food matrixes without considerable  
353 increase of costs. Alterations in color, anthocyanin, total polyphenolic, radical scavenging activity and reducing  
354 power showed a major effect caused by drying conditions (time and temperature), and their levels were dependent

355 on the particular red fruit. In particular, drying berries at 65 °C during 20 h was the best choice for the conservation  
356 of color, polyphenol content and antioxidant activity while anthocyanin content was similar in berries dried at 50 °C  
357 or 65 °C. It is clear that drying at temperature above 100 °C greatly deteriorates all the berry properties analyzed. It is  
358 remarkable that berries dried at 65 °C could be a great option to produce berry powder in order to improve the  
359 nutritional and sensorial characteristics of the final products. Conventional drying is a much more economical drying  
360 method than lyophilization and shows a considerable increase of total polyphenol content due to depolymerization  
361 of native polyphenols that also leads to increased antioxidant activity with pleasant smell and color.

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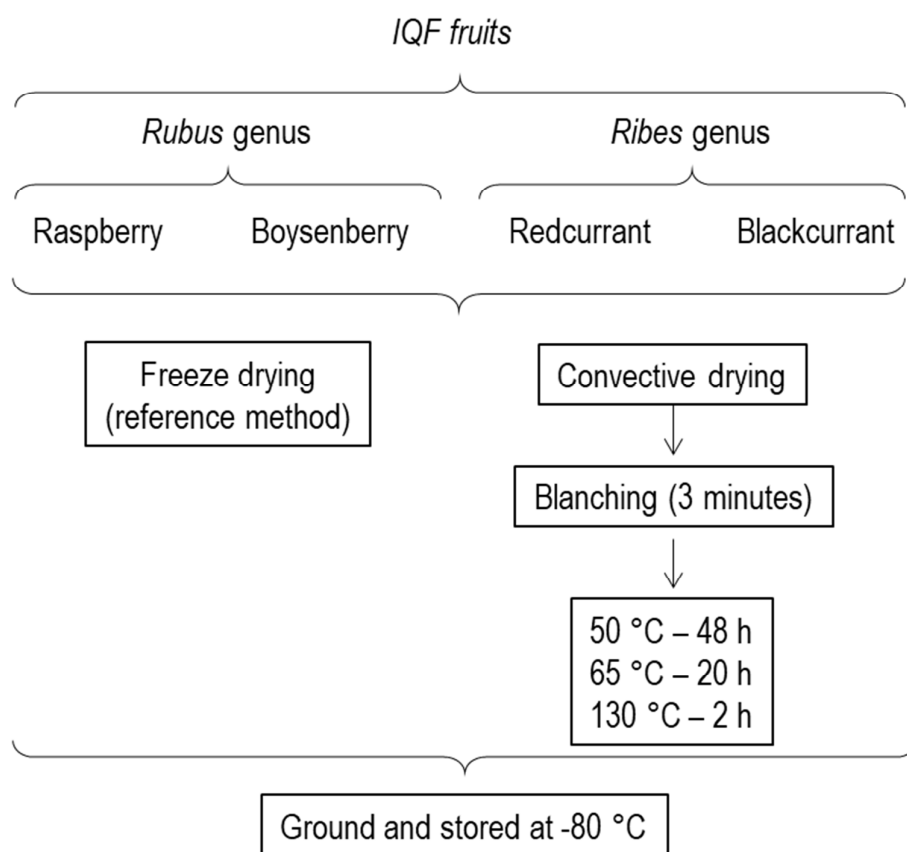


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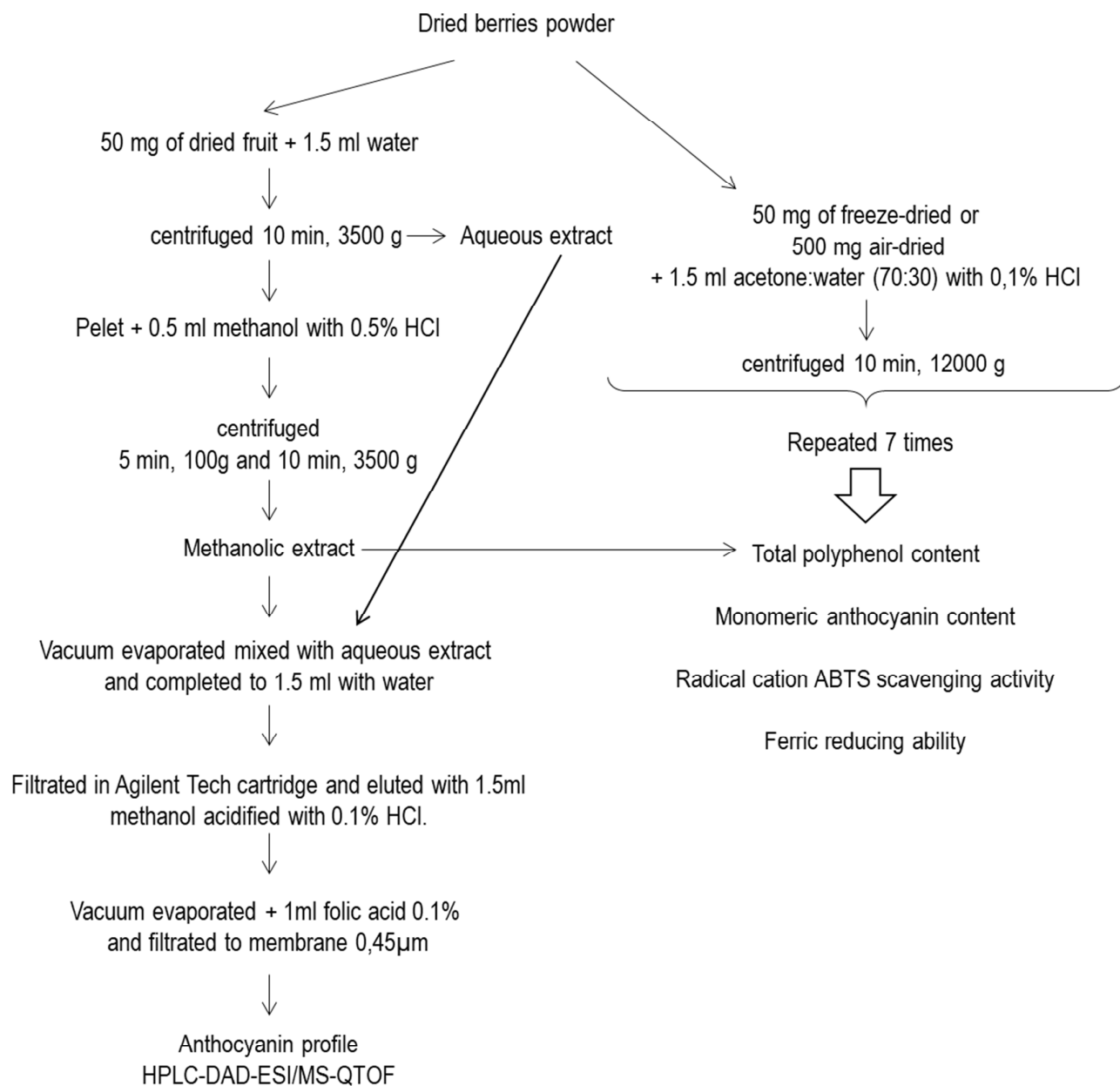
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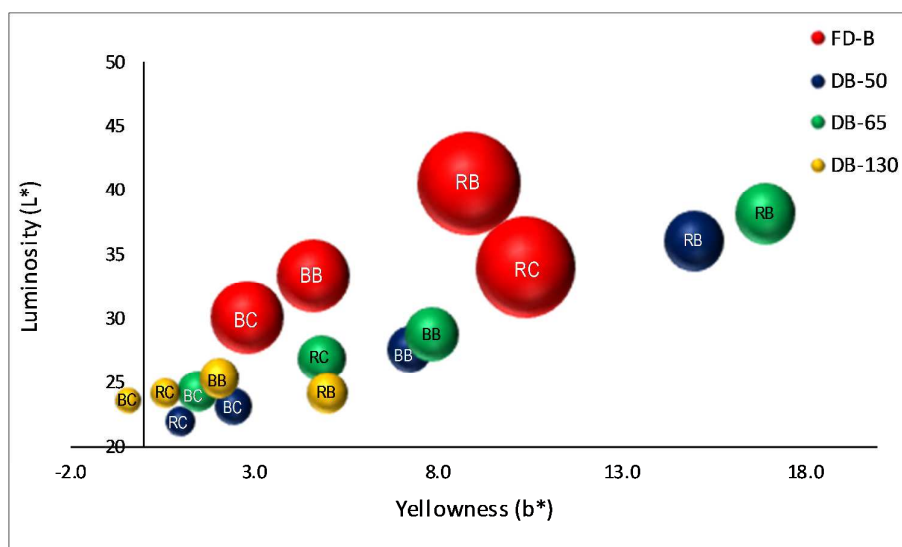
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**Figure 1.-** Berries drying procedure.

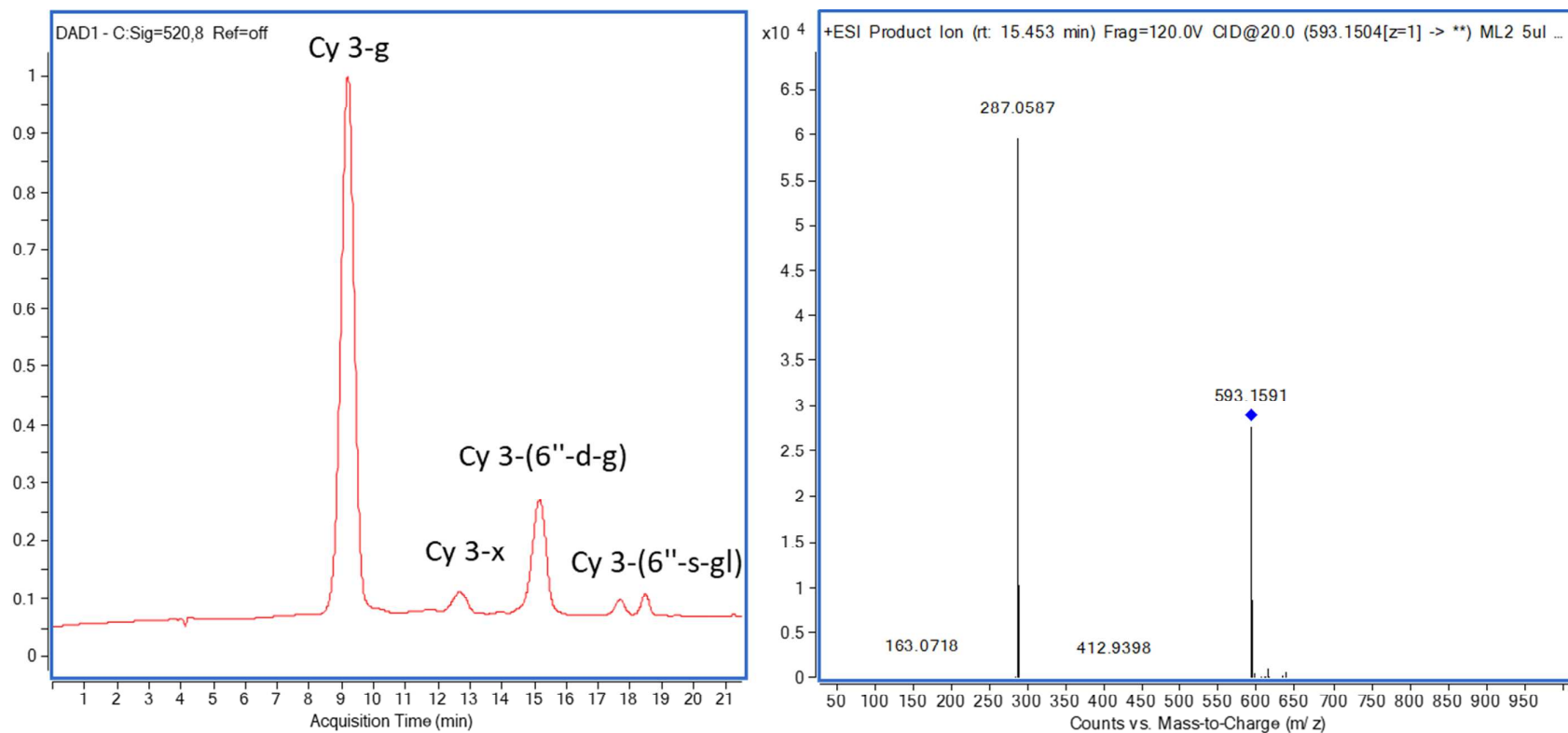


**Figure 2.-** Analytical protocol for anthocyanin profiling, total polyphenol content and antioxidant activity determinations.

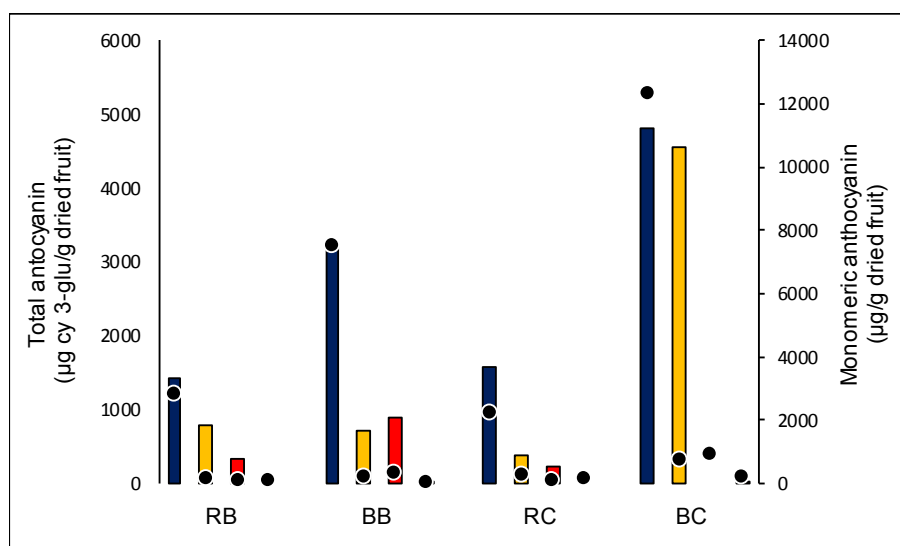


**Figure 3.-** Color parameters of dried berries at different conditions. Bubbles sized are related to redness ( $a^*$ ) value.

Raspberry (RB), boysenberry (BB), redcurrant (RC) and blackcurrant (BC).



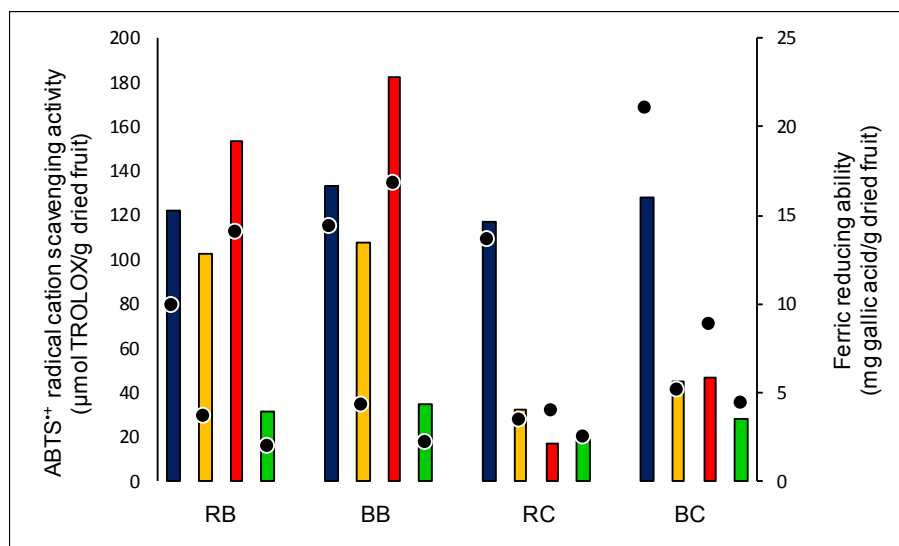
**Figure 4.-** Chromatogram at 520nm corresponding to sample boysenberry (BB) lyophilized (left) and MSMS spectra of the peak eluting at 15.3 min with a  $M^+$  of 593.16 that has been identified as Cyanidin 3-(6''-dioxalyl-glucoside) (right). Cy 3-g , Cyanidin 3-glucoside; Cy 3-x, Cyanidin 3-xyloside; Cy 3-(6''-d-g), Cyanidin 3-(6''-dioxalyl-glucoside) and Cy 3-(6''-s-gl), Cyanidin 3-(6''-succinyl-glucoside).



**Figure 5.-** Total anthocyanin by HPLC-DAD (bars) and monomeric anthocyanin (dots) of freeze-dried berries (blue)

and berries dried at 50 °C (yellow), 65 °C (red) and 130 °C (green).

Raspberry (RB), boysenberry (BB), redcurrant (RC) and blackcurrant (BC). \*logarithmic scale



**Figure 6.-** ABTS<sup>•+</sup> radical cation scavenging activity (bars) and ferric-reducing power (dots) of freeze-dried berries (blue) and berries dried at 50 °C (yellow), 65 °C (red) and 130 °C (green).  
Raspberry (RB), boysenberry (BB), redcurrant (RC) and blackcurrant (BC).



**Table 1.- Physicochemical composition of selected berries\***

	Water content	Ash	Protein	Carbohydrate
Raspberry	85.8 ± 0.4 <sup>c</sup>	1.68± 0.03 <sup>a</sup>	4.7 ± 0.1 <sup>d</sup>	7.9± 0.5 <sup>a</sup>
<i>USDA database</i> <sup>*1</sup>	85.75	-	1.20	11.94
Boysenberry	83.0 ± 0.4 <sup>b</sup>	1.64± 0.02 <sup>a</sup>	3.6 ± 0.0 <sup>c</sup>	11.7± 0.4 <sup>b</sup>
<i>USDA database</i> <sup>*1</sup>	88.15	-	1.39	9.61
Redcurrant	82.4 ± 0.8 <sup>b</sup>	2.06± 0.04 <sup>b</sup>	2.8± 0.0 <sup>a</sup>	12.8 ± 0.7 <sup>b</sup>
<i>USDA database</i> <sup>*1</sup>	83.95	-	1.40	13.80
Blackcurrant	75.4 ± 0.3 <sup>a</sup>	2.93± 0.02 <sup>c</sup>	3.1± 0.2 <sup>b</sup>	18.6± 0.1 <sup>c</sup>
<i>USDA database</i> <sup>*1</sup>	81.96	-	1.40	15.38

\*Within the same column, values with different letters indicate significant differences (P<0.05).<sup>\*1</sup>

USDA standard value for reference: <https://ndb.nal.usda.gov/ndb/search/list>

**Table 2.- Water activity of dehydrated fruits\***

Condition/Fruit	Raspberry	Boysenberry	Redcurrant	Blackberry
Freeze-dried	$0.376 \pm 0.002^d$	$0.357 \pm 0.001^c$	$0.242 \pm 0.000^a$	$0.334 \pm 0.001^b$
50 °C	$0.310 \pm 0.002^c$	$0.413 \pm 0.002^d$	$0.411 \pm 0.002^d$	$0.398 \pm 0.002^d$
65 °C	$0.251 \pm 0.001^a$	$0.326 \pm 0.001^b$	$0.342 \pm 0.003^c$	$0.343 \pm 0.002^c$
130 °C	$0.302 \pm 0.001^b$	$0.280 \pm 0.001^a$	$0.291 \pm 0.001^b$	$0.282 \pm 0.001^a$

**Table 3.- Chroma ( $\Delta C_{ab}$ ) and total color difference ( $\Delta E$ ) for fruits dehydrated by convective drying\***

Air-drying temperature	50 °C				65 °C				130 °C			
Fruit sample	RB	BB	RC	BC	RB	BB	RC	BC	RB	BB	RC	BC
$\Delta C_{ab}$	24.3 <sup>e</sup>	10.7 <sup>b</sup>	31.6 <sup>h</sup>	13.0 <sup>c</sup>	24.5 <sup>b</sup>	8.3 <sup>a</sup>	26.3 <sup>f</sup>	12.4 <sup>c</sup>	29.9 <sup>g</sup>	12.1 <sup>c</sup>	31.7 <sup>h</sup>	15.7 <sup>d</sup>
$\Delta E$	24.8 <sup>e</sup>	12.2 <sup>b</sup>	33.8 <sup>g</sup>	14.8 <sup>c</sup>	24.6 <sup>e</sup>	9.5 <sup>a</sup>	27.2 <sup>f</sup>	13.7 <sup>c</sup>	34.1 <sup>g</sup>	14.5 <sup>c</sup>	33.1 <sup>g</sup>	17.0 <sup>d</sup>

\*Within the same row, values with different letters indicate significant differences (P<0.05).

Raspberry (RB), boysenberry (BB), redcurrant (RC) and blackcurrant (BC).

**Table 4.- Anthocyanin profile in lyophilized berries.**

Peak (n)	Rt (min)	Structure	M+	Fragments MS–MS	fruit
1	7.7	Cyanidin-3-sophoroside	611.16	449.09, 287.05	RB
2	7.7	Delphinidin 3-rutinoside	611.16	465.09, 303.04	BC
3	8.3	Cyanidin 3-sophoroside-5-rhamnoside	757.21	433.1, 287.05	RB
4	9.2	Cyanidin 3-glucoside	449.11	287.05	RB, BB
5	9.2	Cyanidin 3-xylosyl-rutinoside	727.2	287.06	RC
6	9.7	Cyanidin 3 -rutinoside	595.16	449.1, 287.05	RC, BC
7	12.8	Cyanidin 3-xyloside	419.05	287.05	BB
8	15.3	Cyanidin 3-(6"-dioxalyl-glucoside)	593.16	287.05	BB
9	18.4	Cyanidin 3-(6"-succinyl-glucoside)	549.12	287.05	BB

Raspberry (RB), boysenberry (BB), redcurrant (RC) and blackcurrant (BC).

**Table 5.- Quantification of anthocyanins in dried fine fruits.**

Drying conditions	Fruit	Cy 3- sophoroside +	Cy 3- rutinoside	Cy 3- glucoside	D 3- rutinoside	Cy 3- xylosyl- rutinoside	Cy 3- xyloside	Cy 3- (6"-dioxalyl- glucoside)	Cy 3- (6"-succinyl- glucoside)
		Cy 3- sophoroside 5-rhamnoside							
FDB	RB	<b>744±60</b>		676±7*					
	BB			<b>3255±57</b>			145±33	157±50	109±28
	RC				427±33	<b>1145±22</b>			
	BC		<b>2512±282</b>		2305±40				
DB -50	RB	<b>420±41</b>		380±29*					
	BB			<b>713±24</b>			27.7±1	15.1±0	
	RC				53±5	<b>341±34</b>			
	BC		<b>2338±64</b>		2227±14				
DB -65	RB	<b>187±14</b>		148±52*					
	BB			<b>882±12</b>			1±0	65±0	23±0
	RC				52±0	<b>229±1</b>			
	BC								
DB -130	RB								
	BB			<b>37±12</b>			12±1	5±0	21±0
	RC								
	BC		<b>8±0</b>		6±0				

FDB: freeze-dried berry, DB-50: air-dried berry at 50 °C, DB65: air-dried berry at 65 °C, DB130: air dried berry at 130 °C, RB: raspberry, BB: boysenberry, RC: redcurrant and BC: blackcurrant.

Mean ± SD (n ≥ 3). Numbers in the table are in bold to show the most abundant anthocyanin in each sample, while white spaces correspond to not quantifiable anthocyanin. Within the same column, value with different letters indicate significant differences (P<0.05). Nd: not detectable.

\*In RB Cy 3-glucoside co-eluted with Cy 3-rutinoside.

**Table 6.- Total polyphenol content in acetone and methanolic extracts of dried berries\***

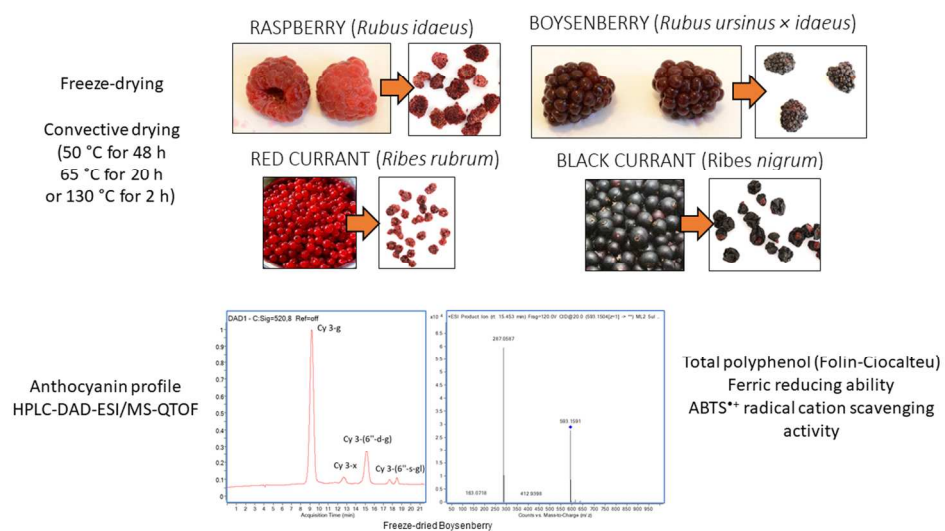
		Total polyphenol (mg gallic acid equivalents/g dried fruit)	
Drying conditions	Fruit	Acidified acetone extract	Acidified methanolic extract
FDB	RB	18.7 <sup>e</sup>	3.2 <sup>b</sup>
	BB	23.8 <sup>h</sup>	5.6 <sup>c</sup>
	RC	21.5 <sup>f</sup>	2.0 <sup>a</sup>
	BC	32.3 <sup>h</sup>	3.9 <sup>b</sup>
DB50	RB	5.1 <sup>c</sup> (-73%)	6.7 <sup>d</sup> (+109%)
	BB	12.5 <sup>d</sup> (-47%)	6.6 <sup>d</sup> (+18%)
	RC	5.8 <sup>c</sup> (-73%)	2.2 <sup>a</sup> (+10%)
	BC	11.6 <sup>d</sup> (-64%)	11.0 <sup>f</sup> (+182%)
DB65	RB	35.2 <sup>i</sup> (+88%)	5.4 <sup>b</sup> (+69%)
	BB	41.6 <sup>k</sup> (+75%)	12.0 <sup>g</sup> (+114%)
	RC	23.1 <sup>g</sup> (+7%)	3.8 <sup>b</sup> (+90%)
	BC	37.2 <sup>j</sup> (+15%)	7.7 <sup>e</sup> (+97%)
DB130	RB	1.4 <sup>a</sup> (-93%)	6.7 <sup>c</sup> (+109%)
	BB	4.0 <sup>b</sup> (-83%)	10.9 <sup>f</sup> (+95%)
	RC	3.6 <sup>b</sup> (-83%)	3.9 <sup>d</sup> (+95%)
	BC	5.8 <sup>c</sup> (-82%)	4.3 <sup>b</sup> (+10%)

FDB: freeze-dried berry, DB-50: air-dried berry at 50 °C, DB65: air-dried berry at 65 °C, DB130: air-dried berry at 130 °C, RB:

raspberry, BB: boysenberry, RC: redcurrant and BC: blackcurrant.

\*Within the same column, values with different letters indicate significant differences (P<0.05)

Values between parentheses indicate percentage of increase or decrease with respect to FDB.



TOC Graphic

338x190mm (96 × 96 DPI)